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Article

A Photo-Enzymatic Cascade to Transform Racemic Alcohols into Enantiomerically Pure Amines

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Abstract: The consecutive photooxidation and reductive amination of various alcohols in a cascade reaction were realized by the combination of a photocatalyst and several enzymes. Whereas the photocatalyst (sodium anthraquinone-2-sulfonate) mediated the light-driven, aerobic oxidation of primary and secondary alcohols, the enzymes (various ω -transaminases) catalyzed the enantio-specific reductive amination of the intermediate aldehydes and ketones. The system worked in a one-pot one-step fashion, whereas the productivity was significantly improved by switching to a one-pot two-step procedure. A wide range of aliphatic and aromatic compounds was transformed into the enantiomerically pure corresponding amines via the photo-enzymatic cascade.

Keywords: photooxidation; cascade; alcohol; reductive amination; ω -transaminase

1. Introduction

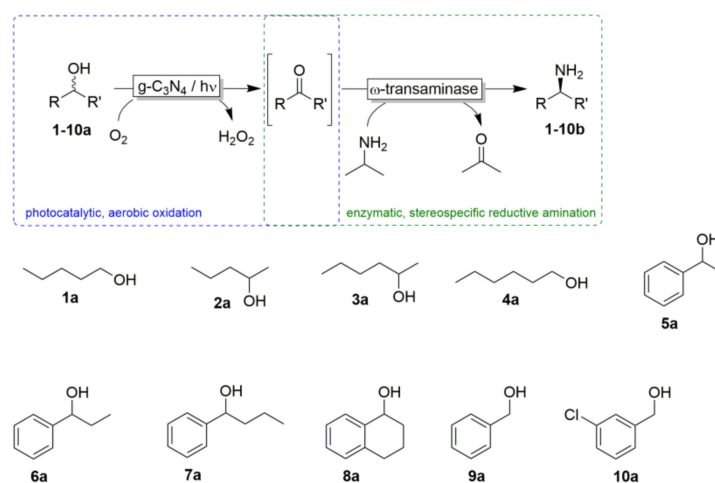
In recent years, significant attention has been devoted to the synthesis of amines [1]. Especially, enantiomerically pure amines are of interest in natural product synthesis, as intermediates in pharmaceutical synthesis, and for a variety of other chemical products [2–4]. Direct amination of alcohols has been studied extensively [1,5]. The main advantage of prompt conversion of alcohols to amines is that both the substrate and the product are in the same oxidation state, and, thus, theoretically the additional use of any redox equivalents is not required [6,7]. Furthermore, many of the required alcohols are already available on an industrial scale, which facilitates the mass application of the technology [8]. On the other hand, most of the currently applied reactions either possess poor chemo-selectivity and require harsh reaction conditions (e.g., the reaction of alcohol with ammonia over various heterogeneous catalysts, such as tungsten, cobalt, nickel, chromium) [5] or have low efficiency and produce toxic intermediates [9]. It is also worth mentioning that the majority of the current methodologies either only accept symmetric substrates or produce racemic products. To synthesize chiral amine moieties, ketones are one of the most commonly used precursors [1].

Various approaches have been reported to selectively oxidize alcohols to the corresponding carbonyl products [10–13]. Compared to transition metal-based approaches [14], photooxidation provides an atom-economic and environmentally benign alternative [10]. Commercially available TiO₂ is used most commonly as a photocatalyst, but the applicability is restricted by its UV absorption [15]. However, efficient oxidation of 1-pentanol has been reported by irradiation up to 480 nm upon doping TiO₂ with Nb₂O₅ [16,17]. To further utilize visible light, numerous other heterogeneous catalysts have been tested [18]. By using graphitic carbon nitride (g-C₃N₄) [19,20] and vanadium-oxide grafted

to numerous carriers [21,22], efficient alcohol oxidation mediated by visible light has been reported. Besides heterogeneous systems, water-soluble sodium anthraquinone-2-sulfonate (SAS) is also known as an effective and inexpensive homogeneous photocatalyst [23–27]. Very recently we found that SAS is also very active for the oxidation of C–H bonds [27] and, therefore, can be implemented in photo-enzymatic cascades for the conversion of a very large substrate scope [28].

Even though photocatalysis provides an attractive approach to produce the carbonyl intermediate, typically, tedious protection/de-protection steps are required for the synthesis of chiral amines. These additional stages prolong the synthesis time and significantly decrease the atom efficiency of the reaction. To aid these shortcomings, biocatalytic approaches have been extensively studied [29–32]. Pyridoxal-5-phosphate (PLP)-dependent ω -transaminases (ω -TAs) provide an elegant way to convert a carbonyl group to an amine moiety at the expense of a sacrificial amine donor [31,33]. ω -TAs have been increasingly applied in industrial chemical synthesis, in particular for the manufacturing of active pharmaceutical ingredients (APIs) [29,34,35]. Other main applications deal with the production of surfactants [36], amino acids [37], and plastic fiber monomers [36,38,39]. To reach an acceptable conversion when L- or D-alanine is used as an amine donor, the reaction equilibrium of the transamination reaction must be shifted towards the product. Thus, one of the most common means to enhance the amine formation is to remove the pyruvate by-product through the addition of another enzyme, such as lactate dehydrogenase (LDH), alanine dehydrogenase (AlaDH), pyruvate decarboxylase (PDC), or an acetolactate synthase (ALS) [32]. On the other hand, the use of alternative amine donors provides a more favorable thermodynamic equilibrium. However, these molecules are either prohibitively expensive [39] or generate a co-product during the transamination that tends to polymerize, thus complicating product isolation and lowering product yield [40,41]. Therefore, it is not surprising that isopropylamine (IPA) has been used as the preferred amine donor, especially on an industrial scale. IPA is inexpensive and provides a much more favorable thermodynamic equilibrium compared to alanine [42], which can be further shifted via simple and selective evaporation of the low boiling acetone co-product [35]. Consequently, IPA was selected as the amine donor in this study.

Recently, numerous reports have been published about the effective conversion of a plethora of alcohols to (chiral) amines in multi-enzyme cascades [43,44]. Inspired by their results and the current advances in the selective photochemical oxidation of alcohols [18], herein we present a photo-enzymatic tandem reaction for the direct conversion of alcohols to (chiral) amines. Employing the consecutive oxidation and reductive amination steps in one pot, a further reduction of the environmental impact of the synthesis is possible (Scheme 1). As time-consuming intermediate isolation and purification steps are omitted, the required amount of organic solvents is minimized, which results in a lower E-factor (*Environmental factor*, defined as the mass of waste per mass of product) [45,46].



Scheme 1. Schematic representation of the photo-enzymatic cascade reaction and the examined substrates.

2. Results and Discussion

In the first set of experiments, we evaluated a range of homogeneous and heterogeneous photocatalysts available in our laboratory. More specifically, water-soluble sodium anthraquinone-2-sulfonate (SAS) and heterogeneous graphitic carbon nitride ($g\text{-C}_3\text{N}_4$), niobium(V)-oxide-doped titanium-dioxide ($\text{TiO}_2\text{-Nb}_2\text{O}_5$), and vanadium oxide (VO) deposited on various carriers, such as alumina ($\text{VO-Al}_2\text{O}_3$), zirconium-dioxide (VO-ZrO_2), and graphitic carbon nitride ($\text{VO-g-C}_3\text{N}_4$), were evaluated. As a model reaction, we chose the oxidation of *rac*-1-phenylethanol (**5a**) to acetophenone (**5b**) (Figure 1). Starting from 25 mM of the starting material, SAS mediated the full conversion of **5a** (TN = 30), while the best heterogeneous catalyst, $g\text{-C}_3\text{N}_4$, produced only 6.2 mM of product with 20% conversion. The discrepancy between conversion and product concentration in the case of SAS is probably due to the oxidative decomposition of the product. Since the other photocatalysts fell back significantly in their performance (Figure 1), we focused our attention on SAS and $g\text{-C}_3\text{N}_4$.

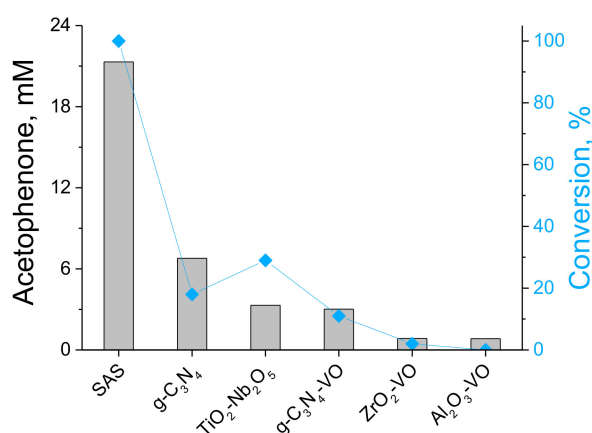


Figure 1. Alcohol oxidation using various photocatalysts. Reaction conditions: 1 mL MilliQ water with [**5a**] = 25 mM, [sodium anthraquinone-2-sulfonate (SAS)] = 0.75 mM, or [heterogeneous photocatalyst] = 10 mg/mL. Reactions were performed for 24 h at 30 °C under atmospheric conditions and irradiated under visible light ($\lambda > 400$ nm). These experiments have been performed as single experiments.

We performed the cascade reaction shown in Scheme 1 using SAS and $g\text{-C}_3\text{N}_4$ in a one-pot one-step fashion, i.e., adding the starting material and all catalysts at the beginning (Figure 2). For the reductive amination of the intermediate ketone, two ω -transaminases were applied: the (*R*)-selective ω -TA from *Aspergillus terreus* (*AT* ω TA) and the (*S*)-selective ω -TA from *Bacillus megaterium* (*BM* ω TA). In the case of $g\text{-C}_3\text{N}_4$, virtually no alcohol conversion was observed. The trace amounts of *rac*-**5c** originated from small acetophenone impurities within the commercial substrate. We attribute this lack of catalytic activity to the absorption of the biocatalysts to the $g\text{-C}_3\text{N}_4$ surface, thereby passivating it for the desired oxidation reaction.

The cascade reaction using SAS, however, produced 2.5 mM (*S*)- and 1.5 mM (*R*)-1-phenylethyl amine in 48 h. Even though enantiomerically pure amines were obtained, the performance of this system fell back behind our expectations. Therefore, we set out to investigate the limitations of the one-pot one-step procedure. Further characterization of the reaction conditions revealed some limitations of the one-pot one-step procedure to be as follows: (i) the catalytic activity decreased steadily in the presence of light and SAS. This may be attributed to an oxidative inactivation and degradation of the biocatalysts by photoexcited SAS and the used light (Figure S9); (ii) we found that the rate of the first step (SAS-mediated oxidation of phenylethanol) was significantly slower in the case of the one-pot one-step procedure. This may be attributed to SAS's activity on the stoichiometric amine donor and the product of the reductive amination step (Figures S11–S13).

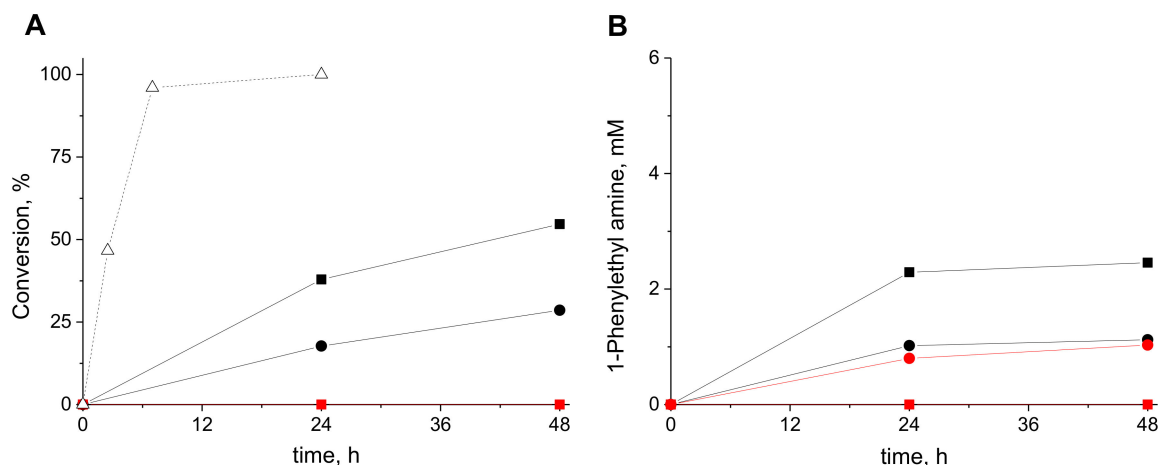


Figure 2. Results of the direct amination of 1-phenylethanol (**5a**) in a one-pot one-step fashion with SAS (—) and g-C₃N₄ (—) photocatalysts, employing *BM*ωTA (*Bacillus megaterium* ωTA ■/■) and *AT*ωTA (*Aspergillus terreus* ωTA ●/●) enzymes. (A) Conversion of **5a**. As a comparison, the conversion of 20 mM **5a** in MilliQ water containing 0.75 mM SAS is also shown (Δ); (B) Production of 1-phenylethyl amine **5c**. Reaction conditions: 1 mL reaction mixture containing sodium phosphate (NaPi) buffer (100 mM, pH 9), [1-phenylethanol] = 15 mM, [SAS] = 0.75 mM, [isopropylamine, IPA] = 1 M, [Pyridoxal-5-phosphate, PLP] = 1 mM, [crude cell extract overexpressed with ω-TA enzyme] = 10 mg/mL. Reaction mixtures were incubated at 30 °C, irradiated with white light (λ > 400 nm).

Therefore, we turned our attention to a one-pot two-step procedure, wherein we first performed the photochemical alcohol oxidation followed by supplementation of the reaction mixture containing the reaction components needed for the reductive amination in a second step. Compared to the one-pot one-step case, the rate of the oxidation increased meaningfully, and the amount of product **5c** doubled with excellent enantiomeric excess (>99%) (Figure 3).

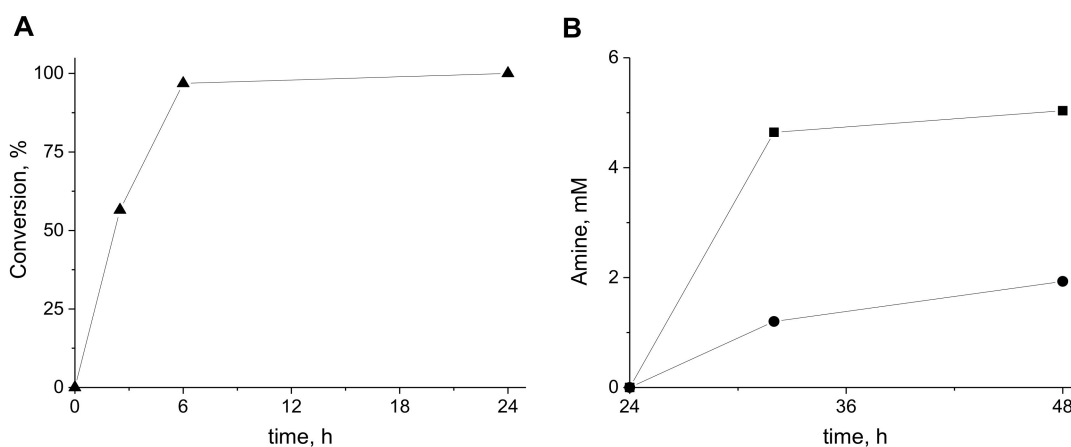


Figure 3. Results of the direct amination of 1-phenylethanol (**5a**) in a one-pot two-step fashion. (A) Conversion of **5a** (▲); (B) Production of **5c** with *BM*ωTA (■) and *AT*ωTA (●) enzymes; Reaction conditions: (A) 1 mL MilliQ water containing [**5a**] = 10 mM, [SAS] = 0.75 mM. Samples were incubated at 30 °C and irradiated under visible light (λ > 400 nm); (B) 0.5 mL pH 9 reaction mixture containing [NaPi] = 50 mM, [IPA] = 1 M, [PLP] = 1 mM, [crude cell extract overexpressed with ω-TA enzyme] = 10 mg/mL. Samples were incubated on a shaking plate at 30 °C at 600 rpm.

To gain more insight into the performance of the ω-transaminases, we varied the enzyme, amine donor, and cofactor concentrations. An increase in the enzyme concentration resulted in a higher initial reaction rate. However, approx. 2.9 mM of product was formed for all cases after 28 h using 1 M of IPA at pH 9 (Figure S23). Using the same concentration of IPA but increasing the pH to 10.2

led to a progressive increase in the yield above 6 mM (Figure S26). These data are in agreement with previous publications, which indicate that the deprotonated form of IPA in aqueous solution might act as the actual amine donor [35,47,48]. Testing different concentrations of IPA at pH 9 (0.5, 1, 2 M) resulted in both higher initial rate and final product concentration (Figure S24), as also the amount of free IPA amino donor in solution increased accordingly. We concluded that the final obtained conversion was determined by the actual concentration of free IPA in solution and the thermodynamic equilibrium constant of the reaction [42], and not by the stability of the biocatalyst. Therefore, the productivity of our system can be increased in the future by process engineering aimed at the selective removal of the acetone co-product. Finally, varying the concentration of the cofactor PLP did not show any noticeable influence on the product formation (Figure S25), which is also in agreement with previous findings. Conversely, an excessively high concentration of PLP is not only inconvenient economically but can also inhibit the ω -TA [49]. Based on the above findings, we chose the following parameters for our next investigation: [with ω -TA enzyme] = 10 mg/mL, [IPA] = 1 M, and [PLP] = 1 mM. The reactions were performed at 30 °C, an optimal temperature for both the photooxidation and reductive amination steps.

To further explore the scope of the one-pot two-step procedure, we investigated a broader scope of starting materials including aliphatic, aromatic, chiral, and non-chiral substrates (Scheme 1 and Table 1).

Table 1. Photocatalytic oxidation of various alcohols.

Substrate	Conversion ^a (%)	Yield ^b (%)
1a ^c	23	74
2a ^c	26	51
3a ^c	29	27
4a ^c	28	27
5a	99	94
6a	>99	74
7a	98	59
8a	88	28
9a	98	88
10a	>99	100

For analytical details see Supplementary Materials. ^a Conversion was determined by GC analysis and calculated as $X = (c_0 - c)/c_0$, where X is the conversion, c_0 is the initial substrate concentration, and c is the final substrate concentration. ^b Yield was determined by GC analysis and calculated as $Y = (c_p - c_{p,0})/(c_0 - c)$, where Y is the yield, $c_{p,0}$ is the initial product concentration, c_p is the final product concentration, c_0 is the initial substrate concentration, and c is the final substrate concentration. ^c 1 mL reaction mixture was used in a 2 mL glass vial and irradiated for 24 h. Reaction conditions: 2 mL MilliQ water in a 4 mL vial, containing [SAS] = 0.75 mM and [substrate] = 10 mM. Samples were incubated at 30 °C and irradiated under visible light ($\lambda > 400$ nm) for 9 h.

Aromatic substrates were oxidized with good (>80%) to excellent (>95%) conversion. However, by increasing the length of the aliphatic side chain from 1-phenylethanol (**5a**) to 1-phenylbutanol (**7a**), the yield decreased significantly. This is likely the result of oxidative decomposition, which is enhanced by the inductive effect of the aliphatic side chain [47]. This is also supported by the results for benzyl alcohol (**9a**) and 3-chlorobenzyl alcohol (**10a**). With the presence of an electron withdrawing group on the aromatic ring, which slightly reduces the activity of the hydroxyl group, the selectivity increased. It is also worth mentioning that besides oxidation, 1,2,3,4-tetrahydro-naphtol (**8a**) reacted with SAS, which was indicated by the absence of the characteristic yellow color of the photocatalyst after irradiation. In contrast to the aromatic substrates, aliphatic compounds demonstrated moderate conversion (>30%). Interestingly, C5 compounds (**1a**, **2a**) showed better yield than C6 compounds (**3a**, **4a**).

Following the photooxidation, the reductive amination step was carried out. The oxidized samples were diluted as such that the reaction mixture contained approx. 5 mM aromatic (except α -tetralone (**8b**) 1 mM) and 0.3–1.5 mM aliphatic substrate. In the reactions, the substrate scopes of five wild-type

ω -TAs were examined, one (*R*)-selective from *Aspergillus terreus* (*AT* ω TA), and four (*S*)-selective ones from *Bacillus megaterium* (*BM* ω TA), *Chromobacterium violaceum* (*CV* ω TA), *Pseudomonas fluorescens* (*PF* ω TA), and *Vibrio fluvialis* (*VF* ω TA). For all ω -TAs tested, alkaline reaction conditions were found to be favorable for the reductive amination reaction (Figures S26–S30).

Amongst the prochiral compounds (Table 2), the entire range of substrates was transformed with excellent stereoselectivity (>99% *ee*). Acetophenone (**5b**) was accepted by all enzymes. In the case of aromatic compounds (**5b–7b**), the conversion decreased progressively with the increasing length of the alkyl chain attached to the aromatic ring. However, *VF* ω TA was an exception, as it similarly mediated the reductive amination of both propiophenone (**6b**) and butyrophenone (**7b**) with 35% conversion. Reductive amination of **8b** was also feasible using *BM* ω TA with excellent stereospecificity, which has been challenging for other systems [47,50]. Amongst the aliphatic substrates, 2-pentanone (**2b**) and 2-hexanal (**4b**) were converted to detectable levels of product only by *AT* ω TA and *VF* ω TA, respectively. However, the modest performances of other enzymes are likely to be the result of the mediocre initial concentration of the substrate, as both compounds have been reported to be efficiently converted to the corresponding chiral amine [47].

Table 2. Reductive amination of ketones.

		2b	3b	5b	6b	7b	8b
<i>AT</i> ω TA	<i>c</i> _{amine} (mM) ^a	0.66 (<i>R</i>)	n.d.	2.8 (<i>R</i>)	n.d.	n.d.	n.d.
	Conversion (%) ^b	36	n.d.	72	n.d.	n.d.	n.d.
	<i>ee</i> (%) ^c	>99	n.d.	>99	n.d.	n.d.	n.d.
<i>BM</i> ω TA	<i>c</i> _{amine} (mM)	n.d.	n.d.	4.93 (<i>S</i>)	10.75 (<i>S</i>)	n.c	0.42 (<i>S</i>)
	Conversion (%)	n.d.	n.d.	95	89	n.d.	20
	<i>ee</i> (%)	n.d.	n.d.	>99	>99%	n.d.	>99%
<i>CV</i> ω TA	<i>c</i> _{amine} (mM)	n.d.	n.d.	1.29 (<i>S</i>)	n.d.	n.d.	n.d.
	Conversion (%)	n.d.	n.d.	34	n.d.	n.d.	n.d.
	<i>ee</i> (%)	n.d.	n.d.	>99	n.d.	n.d.	n.d.
<i>PF</i> ω TA	<i>c</i> _{amine} (mM)	n.d.	n.d.	1.95 (<i>S</i>)	2.40	n.d.	n.d.
	Conversion (%)	n.d.	n.d.	43	35	n.d.	n.d.
	<i>ee</i> (%)	n.d.	n.d.	>99	>99	n.d.	n.d.
<i>VF</i> ω TA	<i>c</i> _{amine} (mM)	n.d.	2.25 (<i>S</i>)	3.73 (<i>S</i>)	2.44	1.20 (<i>S</i>)	n.d.
	Conversion (%)	n.d.	100	72	35	35	n.d.
	<i>ee</i> (%)	n.d.	>99	>99	>99 %	>99	n.d.

For analytical details, see Supplementary Materials. ^a Amine concentration was determined by GC analysis ^b Conversion was determined by GC analysis and calculated as indicated in Table 1. ^c Enantiomeric excess was determined by GC analysis on a chiral stationary phase (see Supplementary Materials). ^d n.d.: Not determined due to low concentration. Reaction conditions: 0.5 mL reaction mixture in a 2 mL glass vial containing [NaPi] 50 mM, [crude cell extract overexpressed with ω -TA enzyme] = 10 mg/mL, [IPA] = 1 M, [PLP] = 1 mM, and [aromatic ketone] \approx 5 mM or [aliphatic ketone] \approx 0.5 mM.

In the case of aldehydes, excellent conversions were achieved (Table 3). Besides *VF* ω TA, all the enzymes completely converted hexanal (**3b**) to hexylamine (**3c**). However, it is also important to mention that similar to the other aliphatic compounds, the initial substrate concentration was low. Both benzaldehyde (**9b**) and 3-chlorobenzaldehyde (**10b**) were fully converted to the corresponding amine.

Table 3. Reductive amination of aldehydes.

		4b	9b	10b
<i>AT</i> ω TA	<i>c</i> _{amine} (mM) ^a	2.21	6.03	8.20
	Conversion (%) ^b	100	100	95
<i>BM</i> ω TA	<i>c</i> _{amine} (mM)	2.21	6.86	11.97
	Conversion (%)	100	100	98

Table 3. Cont.

		4b	9b	10b
CV ω TA	c _{amine} (mM)	2.21	7.00	12.82
	Conversion (%)	100	100	91
PF ω TA	c _{amine} (mM)	2.21	6.84	11.79
	Conversion (%)	100	100.00	91
VF ω TA	c _{amine} (mM)	n.d. ^c	7.19	11.83
	Conversion (%)	n.d.	100.00	98

For analytical details, see Supplementary Materials. ^a Amine concentration was determined by GC analysis ^b Conversion was determined by GC analysis and calculated as indicated in Table 1. ^c n.d.: Not determined due to low concentration. Reaction conditions: 0.5 mL reaction mixture in a 2 mL glass vial containing [NaPi] = 50 mM, [crude cell extract overexpressed with ω -TA enzyme] = 10 mg/mL, [IPA] = 1 M, [PLP] = 1 mM, and [aromatic ketone] \approx 5 mM or [aliphatic ketone] \approx 0.5 mM.

3. Materials and Methods

All chemicals were purchased Sigma-Aldrich (Zwijndrecht, The Netherlands), Fluka (Buchs, Switzerland), Acros (Geel, Belgium) or Alfa-Aesar (Karlsruhe, Germany) with the highest purity available and used without further treatment. The ω -transaminases were prepared via recombinant expression in *Escherichia coli* as described in detail in the Supplementary Materials.

3.1. Reaction Conditions for the One-Pot One-Step Cascade

In a 2 mL glass vial, 1 mL pH 9 reaction mixture was prepared, containing 100 mM sodium phosphate (NaPi), approx. 11 mM 1-phenylethanol (or one of the other alcohols investigated here), 1 mM pyridoxal-5-phosphate (PLP), 0.75 mM sodium anthraquinone-2-sulfonate (SAS), 1 M isopropylamine (IPA), and 10 mg lyophilized crude cell extract overexpressed with (*R*)- or (*S*)-selective ω -transaminase (ω -TA). Samples were irradiated under visible light (Osram Halolux CERAM 205W light bulb; $\lambda > 400$ nm) at 30 °C. The reaction mixture was stirred gently with a magnetic stirrer. At intervals, aliquots were taken, extracted with ethyl acetate, derivatized, and analyzed by gas chromatography (see Supplementary Materials).

3.2. Reaction Conditions for the One-Pot Two-Step Cascade

If not stated otherwise, in a 4 mL vial, 2 mL MilliQ water containing approx. 10 mM substrate and 0.75 mM SAS were irradiated with visible light (Osram Halolux CERAM 205W light bulb; $\lambda > 400$ nm) at 30 °C. The reaction mixture was gently stirred with a magnetic stirrer. At intervals, aliquots were taken, extracted with ethyl acetate, and analyzed by gas chromatography without derivatization (see Supplementary Materials). After the irradiation, 250 μ L reaction mixture was diluted to 500 μ L by using NaPi (100 mM) as such that the final samples contained NaPi (50 mM), IPA (1M), PLP (1 mM), and 10 mg/mL lyophilized whole cells overexpressed with (*R*)- or (*S*)-selective ω -TA. The pH of the system was adjusted to pH 9. Samples were incubated on a shaking plate at 30 °C with 600 rpm shaking speed. At intervals, aliquots were taken, extracted with ethyl acetate, derivatized, and analyzed by gas chromatography (see Supplementary Materials).

3.3. Derivatization of GC Samples

Samples (75 μ L) were extracted with ethyl acetate (150 μ L) dried (MgSO₄, 5 mg) and reacted with acetic anhydride (10 μ L) in the presence of 4-(*N,N*-dimethylamino)pyridine (5 mg) at 45 °C for 45 min. The reaction was quenched with water (75 μ L) and the samples were dried (MgSO₄, 5 mg) again.

4. Conclusions

In summary, a reaction sequence of consecutive photooxidation and reductive amination of various alcohols in one pot is reported. The tandem reaction with sodium anthraquinone-2-sulfonate

(SAS) as a photocatalyst was also feasible in a one-pot one-step approach; however, the yield was significantly inferior to the one-pot two-step case. The system was tested on a plethora of aromatic and aliphatic substrates. Even though all of the compounds were converted, the photooxidation of aromatic substrates proceeded much faster and with a higher yield. In the second step of the cascade reaction, recombinant transaminases originating from *Aspergillus terreus* (AT ω TA), *Bacillus megaterium* (BM ω TA), *Chromobacterium violaceum* (CV ω TA), *Pseudomonas fluorescens* (PF ω TA), and *Vibrio fluvialis* (VF ω TA) were used for the reductive amination. All of the non-chiral substrates were converted with high conversion and the prochiral substrates with excellent enantiomeric excess.

One issue en route to the preparative application is the inactivation of the biocatalysts by the photoexcited photocatalysts. This may be overcome by special separation of the photocatalytic oxidation step from the reductive amination step. For this, for example, immobilized catalysts in a flow chemistry setup may be a doable approach.

Supplementary Materials: The following are available online at <http://www.mdpi.com/2073-4344/9/4/305/s1>. Figure S1: SDS-Page for the expression of the ω TAs; Figure S2: FT-IR spectrum of SAS; Figure S3: FT-IR spectrum of g-C3N4; Figure S4: XRD spectrum of g-C3N4; Figure S5: FT-IR spectrum of g-C3N4; Figure S6: XRD spectrum of g-C3N4-VO; Figure S7: Effect of evaporation on the acetophenone concentration during the photo-oxidation of 1-phenylethanol; Figure S8: Control reaction of the photooxidation; Figure S9: Effect of the circumstances of the photooxidation on the efficiency of the reductive amination; Figure S10: Control reaction of the reductive amination; Figure S11: ¹H NMR spectrum of 1-phenylethyl amine; Figure S12: ¹H NMR spectrum of oxidation of 1-phenylethylamine; Figure S13: ¹H NMR spectrum of oxidation of isopropylamine; Figure S14: Effect of light intensity on the photocatalytic activity of SAS; Figure S15: Effect of light composition on the photocatalytic activity of SAS; Figure S16: Effect of the reaction atmosphere on the photocatalytic activity of SAS; Figure S17: Effect of the amount of photocatalyst on the photocatalytic activity of SAS; Figure S18: Effect of pH on the photocatalytic activity of SAS; Figure S19: Effect of light composition on the photocatalytic activity of g-C3N4; Figure S20: Effect of the reaction atmosphere on the photocatalytic activity of g-C3N4; Figure S21: Effect of the photocatalyst amount on the photocatalytic activity of g-C3N4; Figure S22: Effect of the pH on the photocatalytic activity of g-C3N4; Figure S23: Effect of the enzyme concentration on the yield of the reductive amination; Figure S24: Effect of the amine donor concentration on the conversion of the reductive amination; Figure S25: Effect of the cofactor concentration on the yield of the reductive amination; Figure S26: pH dependency of AT ω -TA; Figure S27: pH dependency of BM ω TA; Figure S28: pH dependency of VF ω TA; Figure S29: pH dependency of PF ω TA; Figure S30: pH dependency of CV ω TA; Figure S31: Representative GC chromatogram of SAS catalyzed photooxidation of 1-pentanol; Figure S32: Representative GC chromatogram of SAS catalyzed photooxidation of 2-pentanol; Figure S33: Representative GC chromatogram of SAS catalyzed photooxidation of 1-hexanol; Figure S34: Representative GC chromatogram of SAS catalyzed photooxidation of 2-hexanol; Figure S35: Representative GC chromatogram of SAS catalyzed photooxidation of 1-phenylethanol; Figure S36: Representative GC chromatogram of SAS catalyzed photooxidation of 1-phenylpropanol; Figure S37: Representative GC chromatogram of SAS catalyzed photooxidation of 1-phenylbutanol; Figure S38: Representative GC chromatogram of SAS catalyzed photooxidation of 1,2,3,4-tetrahydro-1-naphthol; Figure S39: Representative GC chromatogram of SAS catalyzed photooxidation of benzyl alcohol; Figure S40: Representative GC chromatogram of SAS catalyzed photooxidation of 3-chlorobenzyl alcohol; Figure S41: Representative GC chromatogram of derivatized n-pentylamine; Figure S42: Representative GC chromatogram of derivatized 2-aminopentane; Figure S43: Representative GC chromatogram of derivatized 1-hexanol, hexanal and 1-hexylamine mixture; Figure S44: Representative GC chromatogram of derivatized 2-aminohexane; Figure S45: Representative GC chromatogram of derivatized 1-phenylethanol, acetophenone and 1-phenylethyl amine; Figure S46: Representative GC chromatogram of derivatized 1-phenylpropanol, propiophenone (R) and (S) 1-phenylpropyl amine; Figure S47: Representative GC chromatogram of derivatized 1,2,3,4-tetrahydro-1-naphthol, α -tetralone, (R) and (S) 1,2,3,4-tetrahydro-1-naphthylamine; Figure S48: Representative GC chromatogram of derivatized benzyl alcohol, benzaldehyde, benzyl amine; Figure S49: Representative GC chromatogram of derivatized 3-chlorobenzyl alcohol, 3-chlorobenzaldehyde, 3-chlorobenzyl amine, Table S1: Used GC systems; Table S2: Details of GC analysis of the alcohol oxidation; Table S3: Details of GC analysis of the reductive amination.

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