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Characterisation of the Impact of Dissolved Organic Matter on Iron,

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Manganese, and Arsenic Mobilisation during Bank Filtration

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9 Abstract

Bank filtration (BF) has been used for many years as an economical technique for providing 10 11 high-quality drinking water. However, under anaerobic conditions, the aquifer release of 12 undesirable metal(loid)s, such as iron manganese, and arsenic, reduces the bank filtrate quality and thus restricts the application of this technique. This study investigates the impact of the 13 14 organic-matter composition of source water on the mobilisation of Fe, Mn, and As during the anaerobic BF process. A laboratory-scale column study was conducted at a controlled-15 temperature (30±2°C) using different feed water sources. The organic matter characteristics of 16 the feed water were elucidated using excitation-emission spectroscopy techniques integrated 17 with parallel factor framework clustering analysis (PFFCA) model. The experiment was 18 19 performed at redox conditions between 66 mv and -185 mv. Moreover, batch studies were implemented to study the effect of natural organic matter type (humic, fulvic and tyrosine) and 20 concentration on the mobilisation of the selected metal(loids). The laboratory experiments 21 22 demonstrated that the mobilisation of Fe, Mn and As during the BF are varied with the organic water concentration and composition of the source water. The fluorescence results revealed 23 that terrestrial and condensed structure humic compounds are more capable to release Fe into 24 the filtrate water. In contrast, Mn exhibited an equal tendency of mobilisation towards all the 25 26

humic compounds regardless of its origin and structure. However, at a humic concentration

27 higher than 5 mg-C/L, Mn showed more affinity towards lower molecular weight humic compounds. Arsenic was found to be the least impacted by the alteration in the source water 28 organic matter composition; its mobilisation was highly correlated with iron releasing process. 29 30 On the other hand, the biodegradable organic matter at high concentration (>10 mg-C/L) was 31 found to be highly effective to turn the infiltration area into Fe-reducing environment and thereby elevating Fe and As concentrations in the pumped water. In conclusion, this study 32 33 revealed that the DOM composition and concentration of the raw water could play an important role in the mobilisation of metal(loids) during the BF processes. 34

35 Keywords: Bank filtration, Iron, Manganese, Arsenic, Organic Matter

36 1. Introduction

Bank filtration (BF) has been known to be a sustainable and natural treatment technique for 37 38 water supplies worldwide (Eckert et al., 2006). This technique is based on inducing surface water to flow towards a vertical or horizontal well located near a surface water system. The 39 river bank acts as a robust barrier filter to reduce the pollutant concentration. Irmscher et al. 40 (2002) reported that there are more than 100 BF fields in Europe; many of them have been 41 operated for more than 100 years. Recently, BF has been getting increasing attention as a 42 43 desirable technique for treating water supplies in several developing countries with different 44 and extreme environmental conditions (Wahaab et al., 2019).

BF is an effective technique to remove presistent pollutants and produce high drinking water quality. Sandhu et al. (2019) reported a 50% removal of dissolved organic matter (DOM) and 13-99% of micro-organic pollutants for BF fields along the Yamuna River (India). Moreover, BF can also be used as a pre-treatment step to improve the quality of the water and reduce the treatment cost. Haas et al. (2018) revealed that BF could reduce the fouling development in membrane treatment plant and this subsequently will reduce the energy consumption, increase the efficiency and life-time of the membrane, and therefore, reduce the operation cost of theplant.

53 BF is an effective technique to remove persistent pollutants and produce high drinking water 54 quality. Sandhu et al. (2019) reported a 50% removal of dissolved organic matter (DOM) and 13-99% of micro-organic pollutants for BF fields along the Yamuna River (India). Moreover, 55 56 BF can also be used as a pre-treatment step to improve the quality of the water and reduce the 57 treatment cost. Haas et al. (2018) revealed that BF could reduce the fouling development in membrane treatment plants and this subsequently will reduce the energy consumption, increase 58 59 the efficiency and life-time of the membrane, and therefore, reduce the operation cost of the plant. However, other BF processes (e.g., mixing the infiltrate water with the contaminated 60 61 ambient groundwater and reduction of toxic metals from soil into the filtrate water) might also 62 have an adverse effect on the quality of the pumped water. In the last decades, the quality of the River Rhine at Dusseldorf (Germany) was deteriorated and an anaerobic environment was 63 developed in the infiltration area that led to increase the concentrations of Fe and Mn in the 64 pumped water. In the last years, the water quality of the raw water improved; however, the 65 periodic changes of river water quality and hydraulics as a result of climate conditions should 66 67 be considered. Recently, granular activated carbon combined with ozonation and filtration are 68 used as a further treatment of bank filtrate to remove the chemical pollutants (Eckert et al., 69 2006). Thus, it can be concluded that the BF processes and environmental condition of the 70 infiltration area highly determine the quality of the pumped water.

Biodegradation and adsorption are the main mechanisms to remove chemical pollutants during the filtration process (Bertelkamp et al., 2014). Biodegradation is a process in which the microorganisms degrade organic matter using acceptor electrons. Organisms use oxygen as the primary electron acceptor for the organic-matter biodegradation process, followed by nitrate, Mn, and Fe. This process is highly useful to reduce the concentration of organic matter and

76 nitrogen and consequently increase the quality of the pumped water. However, turning the 77 environment into anaerobic, where the insoluble Fe and Mn are the primary electron acceptor, will increase the metals concentrations in the infiltrate water (Massmann et al., 2008). The 78 79 microbial reduction and mobilisation of Fe and Mn are considered as significant sources of elevated toxic substances (e.g., arsenic (As)) in pumped water, which adversely affects human 80 health and degrades bank filtrate quality (Yang et al., 2015). These processes are mainly 81 controlled by environmental conditions along the subsurface flow path, such as the 82 temperature, hydraulic conductivity of the soil, redox potential, metal speciation in the ores, 83 84 and the organic and inorganic compositions of source water (Vega et al., 2017).

DOM is considered as the primary energy source for the metal and metalloids (has intermediate 85 properties of metals and non-metals) mobilisation taking place during the subsurface water 86 87 flow process (Vega et al., 2017). DOM can be subdivided into two major categories namely, 88 biodegradable and non-biodegradable. Biodegradable compounds enrich the biological activity along the flow path and subsequently accelerate the microbial reduction of metal(loid)s into 89 90 the filtrate water (Schittich et al., 2018). Non-biodegradable compounds (i.e., humics) have high electron shuttle capacities and thus could play a dual role in these mobilisation processes. 91 92 First, they could enhance the mobilisation of metal(loid)s biotically by mediating the microbial reduction processes (Brune et al., 2004). Second, they might chemically form aqueous 93 94 complexes with metal(loid)s and thereby increase their concentrations in filtrate water (Liu et 95 al., 2011). On the other hand, the microbial reduction of Fe and Mn might also affect the dynamics and alter the chemical structures of DOM during infiltration. Vega et al. (2017) 96 highlighted that the microbial reduction process is concurrent with the oxidation of labile 97 98 compounds (e.g., phenol) into refractory humic compounds, as well as the destruction of highmolecular-weight organic compounds into smaller compounds that can serve as electron donors 99 for microorganisms during the microbial reduction processes. The dynamics of DOM and its 100

impact on the mobilisation of metal(loid)s during the subsurface flow of water is highly
dependent on the chemical structure of DOM and environmental conditions such as the redox
potential and the temperature.

104 The main objective of this research was to assess the effectiveness of this technique in providing high-quality water in hot-climate environments and to address the problems that 105 restrict the extended application of this technique. The bank filtrate temperature of wells 106 located along the Nile River (Egypt) was 26.4 °C (Ghodeif et al., 2016), whereas, it was 107 reported as 28.6 and 30 °C along the Ganga River and Yamuna River (India) (Sprenger et al., 108 109 2012). Moreover, recent climate models expect a 12.6 °C global average temperature rise by 2050 (R.K. Pachauri et al., 2014). The depletion of oxygen in surface water systems is more 110 likely in arid environments. Hence, there is a higher potential of developing an anaerobic 111 112 environment during the BF processes, which subsequently leads to the enrichment of pumped water in metal(loid)s and degrades the bank filtrate quality. This is one of the major drawbacks 113 that restrict the widespread application of the BF technique in hot-climate regions. Therefore, 114 this research investigated the role of DOM composition in the mobilisation of Fe, Mn, and As 115 during the BF processes under hot-climate conditions. This is an important step to predict and 116 control their concentrations in pumped bank filtrate water. During this research, laboratory-117 scale column and batch studies were conducted using different feed water sources. The organic 118 composition of the feed water was investigated using different fluorescence techniques. 119

120 2. Research Methodology

121 2.1. Soil and feed water characteristics

Manganese iron coated sand (MICS) used in this research was obtained from the groundwater water treatment plant Brucht (Netherlands). The media was sieved through a 3 mm mesh screen, washed gently with water to discard the deposits, and then dried at 70 °C. The size of the used media was 1-3 mm. The physical and chemical properties of the media weredetermined and presented in table S1.

During this study, four different types of waters were fed to the columns to assess the impact of organic matter composition on the mobilisation of Fe, Mn and As during soil passage. The quality parameters of the feed water (pH, conductivity, ammonia, nitrate, total phosphorus, and chlorides) are summarized in table S2. The redox potential of the effluent water was ranged between -66 mv and -185 mv and pH varied between 7.56 and 8.14 during the experimental period.

133 2.2. Column experiment

A laboratory-scale column study was performed to evaluate the role of the organic composition 134 of water in the release of Mn, Fe and As from the soil into the bank filtrate at a high temperature. 135 136 Four PVC columns with 2.1 cm internal diameter and 50 cm height were developed. The columns were packed at the bottom with a support layer of graded gravel with a diameter 3-5 137 mm to a height of 5 cm and then filled with MICS in deionised water to ensure the 138 homogeneous packing. The feed water was introduced to the columns in an up-flow mode 139 (saturated flow), and a variable-speed peristaltic pump was used at the top of the column 140 providing at a constant hydraulic rate (0.5 m/d) to avoid the entrapment of air bubbles in the 141 soil bed. This hydraulic rate has been experienced in many BF fields (Ghodeif et al., 2016; 142 Paufler et al., 2018). The tracer test using NaCl was conducted to estimate the retention time; 143 the empty bed contact time was 16 hours. Sub-oxic conditions were maintained in all columns 144 by continuously degassing the feed water tank with a nitrogen stream to dissipate the air until 145 the dissolved oxygen concentration was lower than 0.2 mg/L. Moreover, the column was fed 146 continuously with water during the whole experiment to prevent the penetration of air into the 147 soil bed. Then, the columns were acclimated using Delft canal water (DCW) for more than 70 148 days. After that, the columns were fed with waters of different organic composition. The first 149

150 column continued to be fed with DCW. The second column was fed with DCW mixed with secondary treated wastewater effluent from Hoek van Holland, the Netherlands (DCWW). The 151 third column was fed with DCW mixed with water-extractable organic matter (WEOM) that 152 had high humic content. The procedures used for WEOM preparation are described in detail in 153 (Abdelrady et al., 2018). The last column was fed with non-chlorinated tap water (NCTW), 154 representing low-organic-matter-content water. The experiment was conducted in a controlled-155 temperature room (at 30 °C). The pore volume (PV) of each column was changed 10–15 times 156 during the whole experiment. Influent and effluent samples were taken regularly (at every PV 157 158 change) to characterise their quality parameters. Physical and chemical parameters (i.e., pH, redox potential, temperature, and dissolved oxygen concentration) were measured 159 continuously to ensure that the experiment was conducted under the desired environmental 160 161 conditions.

162 2.3. Batch experiment

Laboratory-scale batch reactors were employed to determine the impact of the type and 163 concentration of natural organic matter (NOM) on the mobilisation of Fe, Mn, and As from the 164 soil into the filtrate water. Each reactor (0.5 L brown glass bottle containing 100 g MICS) was 165 166 ripened with 400 mL of DCW and the ripening process was continued for 70 days. The feed 167 water was degassed with a nitrogen stream to dissipate the air. Then, the reactors were sealed 168 tightly and placed on a reciprocal shaker at 100 rpm in a controlled-temperature room (30 °C). 169 The renewal process was conducted inside an anaerobic chamber to ensure that the experiment was conducted under anaerobic conditions. After ripening, three NOM types (humic, fulvic, 170 and tyrosine) were spiked at four different concentrations (5, 10, 15, and 20 mg-C/L) in the 171 172 reactors (one type and one concentration in each), and the experiment was conducted for 30 days at the same experimental conditions. This residence time was recommended by several 173 previous studies to remove chemical and biological contaminants from the raw water during 174

the BF processes (Ray et al., 2011). The humic acid and tyrosine were purchased as powders from Sigma-Aldrich (Netherlands) and the powdered fulvic acid was bought from the International Humic Substances Society. Control (DCW without sand) reactors were operated under the same environmental conditions.

179 2.4. Analytical methods

Influent and effluent samples were collected and analysed directly to avoid the degradation of the organic matter. The samples were first filtered using 0.45 µm filtration (Whatman, Dassel, Germany). The Fe and Mn concentrations were determined using inductively coupled plasma– mass spectrometry instrument (Xseries II Thermo Scientific, Bermen, Germany). The arsenic concentration was analysed using a graphite furnace atomic absorption spectrophotometer (Solaar MQZe, Thermo Electron Co). The limit of quantitation (LOQ) of Fe, Mn and As was 10, 10 and 2 µg/L, respectively.

The organic matter content of the feed waters was measured as dissolved organic carbon (DOC, in mg-C/L) using the combustion technique with a total organic carbon analyser (TOC-VCPN (TN), Shimadzu, Japan). The organic composition of the influent and effluent water was determined using the fluorescence excitation–emission technique (F-EEM). A Fluoromax-3 spectrofluorometer (HORIBA Jobin Yvon, Edison, NJ, USA) was used to determine the fluorescence intensity (FI) of the water samples at excitation wavelengths (λ ex) of 240–452 nm (interval = 4 nm) and emission wavelengths (λ em) of 290–500 nm (interval = 2 nm).

194 2.5. Fluorescence indices

195 Fluorescence indices, including the humification index (HIX), the fluorescence index (FIX), 196 and the biological index (BIX), were estimated based on the F-EEM measurements. HIX is 197 used as a marker of the humification (polycondensation) degree of the water, and higher HIX values refer to higher humic content (lower H:C) (Gabor et al., 2014). This parameter wasestimated following the equation proposed by (Ohno, 2002):

200
$$HIX = \frac{\sum FI_{\lambda em (434-480 \text{ nm})}}{\sum FI_{\lambda em (434-480 \text{ nm})} + \sum FI_{\lambda em (300-344 \text{ nm})}} \quad at \ \lambda ex \ (254 \ nm)$$
(1)

FIX is used widely to identify the origin of DOM in water. It is estimated following the method developed by Hansen et al. (2016) (see Equation 2). Its values range between 1.2 and 1.8, with higher values for terrestrial DOM and lower values for microbial DOM origin (Gabor et al., 2014).

205
$$FIX = \frac{FI_{\lambda em} (450)}{FI_{\lambda em} (500)} \quad at \ \lambda ex \ (370 \ nm) \tag{2}$$

BIX is used as an indicator of the contribution of recently produced and microbial-derived organic compounds to the fluorescence DOM pool. It was determined following the method of following the method of Huguet et al. (2009) (Equation 3).

$$BIX = \frac{FI_{\lambda \text{em}}(380)}{FI_{\lambda \text{em}}(430)} \quad at \ \lambda ex \ (310 \ nm)$$
(3)

210 2.6. Fluorescence modelling

Although F-EEM is a qualitative technique, it is still effective in quantifying the characteristics of DOM. A parallel factor framework-clustering analysis (PFFCA) model was recently stipulated by Qian et al. (2017) for decomposing the F-EEM dataset into major fluorescence components representing different NOM compounds. Briefly, PFFCA decomposes the F-EEM datasets into several factors (7–13). Afterwards, the highly correlated factors are clustered into one component representing organic matter with the same fluorescence characteristics.

To ensure the quality of the data and the robustness of the fluorescence dataset interpretation, parallel factor (PARAFAC) analysis was used as well to decompose the fluorescence dataset into major components and to assess the contribution of each component to the full 220 fluorescence spectrum. The development and validation of the model developing were described in details in Murphy et al. (2013). Split-half validation technique was used to validate 221 the fluorescence model. This technique is based on the comparison of multiple models 222 223 generated by splitting the dataset. In this research, the dataset was divided into four groups, and the samples were assigned alternately into the groups. Then, the groups were assembled 224 into six combined groups; each combined group contains half of the dataset. After that, the 225 PARAFAC test was applied for each combined group and the produced models were compared 226 (Harshman et al., 1994). The validation process was implemented using drEEM MATLAB 227 228 toolboxes developed by Murphy et al. (2013).

The redox index (RI) is used as an indicator of the reducibility of quinone-like moieties in the DOM of raw water. This parameter was calculated following the definition of Miller et al. (2006), by dividing the sum of the FI of the reduced quinone-like moieties to the total of the (reduced and oxidised) quinone-like moieties of the fluorescence components, with values close to 1 indicating a higher reducing capacity of the feed water (Gabor et al., 2014).

234 2.7. Statistical analysis

The relationship between Fe, Mn, and As concentrations in the effluent waters and the fluorescence organic characteristics of the feed water were explored using Pearson correlation (r) and nonparametric Spearman rank correlation (ρ) analysis. The strength of the correlation was considered high when the correlation value (ρ) was higher than 0.7 and moderate when it ranged between 0.7 and 0.5 (Hinkle et al., 1988). The relationship was considered significant if the criterion level of significance (p) was lower than 0.05.

241 3. Results

3.1. Impact of DOM composition on metal(loids) mobilisation (column experiment)

243

3.1.1.Fluorescence components

A F-EEM dataset of 80 water samples collected from the influents and effluents of the columns 244 was used to develop and validate a PFFCA-EEM model. The dataset was decomposed into 11 245 factors and afterwards clustered into four main components (FC1-FC4). To ensure the 246 reliability and interpretability of the F-EEM dataset, it was also analysed using the PARAFAC 247 technique. A model with four fluorescence components (PC1-PC4) was successfully split-half-248 249 validated and explained 99.7% of the data variability. The footprints of the PFFCA components and their corresponding PARAFAC components is presented in Figure 1 and Figure S1. A 250 251 comparison between the spectral characteristics of the recognised components and those 252 reported in earlier studies was conducted using OpenFluor online database (Table S1). Component FC1 exhibited maximum wavelengths of $\lambda ex = 332$ nm and $\lambda em = 460$ nm and 253



Figure 1. Contour plots of the four PFFCA components (PF1-PF4) identified from the complete measured F-EEMs dataset for the influents and effluents water of column experiment.

therefore it could be allocated to a terrestrial humic component (Coble, 1996). Singh et al. (2013) reported that this component is ubiquitous in a reduced environment and holds reduced quinone-like moieties in its structure. Component FC2 showed a maximum peak at $\lambda ex = 310$ 257 nm and $\lambda em = 410$ nm, and therefore it was as attributed to a combination of terrestrial fulvic acid and microbial humic compounds (Gonçalves-Araujo et al., 2016; Li et al., 2016). 258 Component FC4 displayed a maximum peak at $\lambda ex = 300$ nm and $\lambda em = 410$ nm. This 259 component is frequently associated with marine and/or microbial humic fluorophores and was 260 recently connected to humic/fulvic compounds of agricultural origin (Baghoth et al., 2011). 261 According to Singh et al. (2013) component FC2 and component FC4 contain oxidised 262 263 moieties in their structures. Based on these definitions, the RI was calculated as the ratio of the first component (FC1) to the sum of the three humic components (FC1, FC2, and FC4). 264 265 Component FC3 exhibited maximum fluorescence at the shortest wavelengths ($\lambda ex = 240$ nm, 268 nm) and ($\lambda em = 308$ nm), which corresponds to a protein-like fluorophore (tyrosine and 266 tryptophan-like fluorophores) (Wünsch et al., 2017). 267

268

3.1.2. Feed water DOM characteristics

Four feed waters (DCW, DCWW, WEOM, and NCTW) with different organic compositions were used in the experiments. Table 1 shows that WEOM had the highest concentration of organic matter (DOC ranged between 15.8 and 19.3 mg-C/L) and the NCTW influent had the lowest one (DOC varied from 1.98 to 4.1 mg/L). However, the DOC levels of DCW and DCWW were 10.8–13.6 mg-C/L and 10.2–13.7 mg/L, respectively.

274 The fluorescence indices were used to elucidate the DOM characteristics of the feed waters (Table 1). The results showed that DCW and DCWW had relatively similar fluorescence DOM 275 characteristics. The HIX, FIX, and BIX of the DCW feed water were 0.85, 1.25, and 0.54, 276 respectively, whereas they were 0.83, 1.26, and 0.61 for DCWW. WEOM had the highest 277 humic content (HIX = 0.9) and the lowest microbial-derived (autochthonous) DOM content 278 (BIX = 0.47), and its ratio of terrestrial humic DOM to microbial DOM (FIX) was 1.08. By 279 280 contrast, NCTW retained a lower humic content (HIX = 0.62) and higher fresh DOM (BIX = 0.73). There were no notable changes in the fluorescence indexes values for DCW, DCWW, 281

and WEOM feed water during the filtration process. However, an increase in the terrestrial humic content (HIX = 0.79, FIX = 1.2) for the NCTW effluent was recorded.

The PFFCA-EEM technique was used to gain insight into the DOM characteristics of the 284 influent waters. Table 1 shows the average FI of PFFCA-EEM components for all of the feed-285 water types. The results demonstrated that the WEOM influent had a considerably higher 286 287 concentration of humic compounds than the other influents. Terrestrial humic (FC1) was present in a proportion (45–49%) of the fluorescence spectrum of WEOM feed water; however, 288 it only represents 43-46%, 41-42%, and 33-34% of the DCW, DCWW, and NCTW 289 290 fluorescence spectrum, respectively. The average FIs of microbial humic FC2 and humic-like from agricultural sources, FC4, were 2.36±0.42 and 0.24±0.04 RU, while their values for 291 NCTW were 0.2±0.09 and 0.1±0.03 RU, respectively. The humic-like components (FC1, FC2, 292 293 and FC4) exhibited a decreasing behaviour during the filtration; the FI of the humic components decreased by 8–26% for DCW, 11–24% for DCWW, and 20–32% for WEOM 294 influent water. An exception was the terrestrial and processed humic compounds of the NCTW 295 influent, which followed an increasing behaviour during the filtration process. On average, the 296 FI of FC1 and FC2 increased by a factor of two and three, respectively, during the filtration 297 298 process. The enrichment of humic compounds in the effluent water might be attributed to: (i) 299 the dissolution of soil organic compounds into the filtrate water, (ii) the presence of micro-300 organisms that are able to bio-transform the labile organic matter into more condensed and 301 refractory compounds (Abdelrady et al., 2018). DCWW and WEOM had the highest concentration of protein-like fluorescent compounds. The average FI of FC3 was 0.47±0.19, 302 0.52±0.14, 0.15±0.08, and 0.65±0.23 RU for DCW, DCWW, NCTW, and WEOM influents, 303 304 respectively. For all feed-water types, the protein-like component FC3 exhibited a decreasing trend during the filtration process; its removal ranged between 67% and 80%, which is in 305

agreement with Abdelrady et al. (2019), who observed an average removal of 80% of labileorganic compounds during the filtration process.

Parameter	Units	DCW	NCTW	DCWW	WEOM	
DOC	mg-C/L	11.64±3.51	3.98±0.73	11.73±1.06	18.23±1.6	
HIX		0.85±0.02	0.62±0.04	0.83±0.06	0.9±0.02	
FIX		1.25±0.07	1.35±0.02	1.26±0.14	1.08±0.13	
BIX		0.54±0.11	0.73±0.04	0.61±0.17	0.47±0.11	
FC1	RU	1.48±0.22	1.44±0.24	0.23±0.09	2.63±0.64	
FC2	RU	1.10±0.19	1.53±0.21	0.20±0.09	2.36±0.42	
FC3	RU	0.47±0.19	0.52±0.14	0.15 ± 0.08	0.65±0.23	
FC4	RU	0.20±0.03	0.15±0.06	0.10±0.03	0.24±0.04	

Table 1. The organic characteristics of the feed waters

309

3.1.3.Iron, manganese and arsenic mobilisation

Figure 2 shows the concentrations of Fe, Mn, and As in the effluents of the columns fed with 310 different water sources. The figure shows that the mobilisation of Mn was much higher than 311 that of Fe and As during the filtration processes. The concentration of Fe, Mn, and As for the 312 four different water types were 10–20 μ g/L, 1500–3900 μ g/L, and >2–7.1 μ g/L, respectively. 313 The concentrations of the tested metal(loid)s in the effluents were found to be dependent on 314 the feed-water source. The WEOM feed water exhibited the highest capacity to release Fe, Mn, 315 316 and As from their solid forms into the filtrate water. The mean concentration of Fe, Mn, and As in the WEOM effluent were 18.28±1.5 µg/L, 3590±185.3 µg/L, and 5.48±1.2 µg/L, 317 respectively. The DCW and DCWW effluents showed a lower capacity of releasing these 318 metal(loid)s during the filtration process. The Fe, Mn, and As concentrations of the DCW 319



Figure 2. Fe (a), Mn (b), and As (c) concentrations in the effluents of columns operated using different water sources (DCW, DCWW, WEOM, NCTW) (anaerobic, 30°C, HRL=0.5 m/d)

effluent were 11.9–15.1 µg/L, 3100–3600 µg/L, and 3.7–5.1 µg/L, respectively, and those of 320 the DCWW effluent were 12.1–13.9 μ g/L, 3000–3600 μ g/L, and 2.9–4.7 μ g/L, 321 correspondingly. By contrast, NCTW exhibited the lowest capability of releasing the assessed 322 323 metal(loid)s, with Fe and Mn concentrations of 10.98±0.9 µg/L and 1990±384.3 µg/L, respectively. Moreover, the As concentration was always below the detection limit ($\langle 2 \mu g/L \rangle$). 324 A strong correlation ($\rho = 0.77$) was observed between Fe and As concentrations in the effluent 325 waters. By contrast, a weak correlation ($\rho = 0.35$) was detected between the As and Mn 326 concentrations. 327

328 *3.1.4.Relationship between DOM composition and metal(loid) mobilisation*

Figure 3 illustrates that the release of metal(loid)s (Fe, Mn, and As) during the filtration process is highly dependent on the DOM concentration of the feed water. The feed DOM concentration was highly correlated with Fe (ρ =0.89) and Mn (ρ =0.87) concentrations in the effluent waters,



Figure 3. Impact of DOC concentration (mg-C/L) of different water sources (DCW, DCWW, WEOM, NCTW) on Fe (a), Mn (b), and As (c) mobilisation during column experiment (anaerobic, 30°C)

whereas a moderate positive relationship was observed between DOM concentration and effluent As concentrations ($\rho = 0.63$).

334 The fluorescence indexes results illustrate that Fe, Mn, and As release during the anaerobic filtration process is much more dependent on the humic content of the feed water than its 335 autochthonous microbial DOM concentration (Table 2). The correlation between HIX and the 336 337 effluent Fe concentration was observed to be $\rho = 0.87$, whereas it was recorded as $\rho = 0.82$ for the Mn concentration. However, a moderate correlation was observed between HIX and the 338 effluent As concentration ($\rho = 0.51$). On the other hand, the mobilisation of Fe, Mn, and As 339 was observed to be in a negative relationship with the microbial and terrestrial DOM ratio, and 340 the correlation between the FIX of the feed waters and the released metal(loid)s was $\rho = -0.71$ 341 for Fe, $\rho = -0.64$ for Mn, and $\rho = -0.39$ for As. 342

343 The correlation between the released metal(loid) concentrations and the FI of the fluorescence components of the feed waters was estimated and is presented in Table 2 and figures S2-S4. A 344 345 strong positive correlation was observed between the Fe concentration and the FI of the humic components (FC1, FC2, and FC4) in the feed waters. The Mn mobilisation, in contrast, 346 exhibited a strong correlation with the FI of humic components from different sources; the 347 correlations between the Mn concentration and the FI of the PFFCA humic components (FC1, 348 FC2, and FC4) were 0.7, 0.82, and 0.73, respectively. Conversely, the As concentration in the 349 350 column effluent water was found to be in a moderate correlation with microbial humic component (FC2) and in weak correlations with the other humic components (FC1 and FC4). 351 Furthermore, the RI demonstrated a moderate correlation with Fe mobilisation and a weak 352 correlation with Mn and As mobilisation. This infers a role for condensed-structure humic 353 compounds in the mobilisation of Fe into filtrate water. On the other hand, labile fluorescence 354

compounds (FC3) were found to have a greater influence on the mobilisation of Mn than on

those of Fe and As.

		DOC	FC1	FC2	FC3	FC4	RI	HIX	FIX	BIX
Fe	spearman Correlation (ρ)	0.89**	.82**	.88**	.6**	.68**	.64**	.87**	71**	76**
	Pearson correlation (r)	0.83**	.83**	.89**	.58**	.68**	.60**	.77**	71**	71**
	n	40	40	40	40	40	40	40	40	40
Mn	spearman Correlation (ρ)	0.87**	.70**	.82**	.77**	.73**	.33**	.82**	64**	85**
	Pearson correlation (r)	0.89**	.77**	.84**	.75**	.67**	.21**	.85**	59**	72**
	n	40	40	40	40	40	40	40	40	40
As	spearman Correlation (ρ)	0.63**	.32	.57**	.05	.42*	.37*	.51**	39*	57**
	Pearson correlation (r)	0.68**	.48**	.64**	.08	.47**	.41*	.52**	51**	58**
	п	30	30	30	30	30	30	30	30	30

357	Table 2. Correlations between	the tested metal	(loid)s and the	fluorescence	characteristics o	f the	feed waters
557	Tuble 2. Conclutions between	ne restea metal	iona is and the	juantescence	character istics of	,	jeeu muters

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***P*-value is less than 0.001

359 *P-value is less than 0.05

360 3.2. Impact of NOM concentrations on metal(loid) mobilisation (batch study)

The effect of NOM (humic, fulvic, and tyrosine) concentrations on the release of metal(loid)s 361 (Fe, Mn, and As) from the soil into the filtrate water under anaerobic conditions was studied in 362 established batch reactors. The fluorescence characteristics of the injected NOM is presented 363 in Figure S5. The experimental results illustrated that the Fe mobilisation increased steadily 364 with humic and fulvic concentrations (Figure 4.a). Fe exhibited relatively higher affinity with 365 humic (high molecular weight) compounds compared to that with fulvic ones (low molecular 366 weight). The concentration of soluble Fe reached 162±19 μ g/L when the humic concentration 367 was 20 mg-C/L, whereas it was $114\pm28 \mu$ g/L at the same concentration of fulvic acid. On the 368 other hand, tyrosine exhibited lower ability to release Fe at low concentrations; the 369 concentrations of Fe in the effluent water in the batch reactors injected with 5 and 10 mg-C/L 370

of tyrosine were 30 ± 17 and $32\pm11 \ \mu g/L$, respectively. However, an immense increase in Fe concentration was observed for the batch reactors injected with 15 mg-C/L and 20 mg-C/L of tyrosine, where the soluble Fe concentrations were $290\pm47 \ \mu g/L$ and $340\pm24 \ \mu g/L$, respectively.

Figure 4.b illustrates the impact of NOM type and concentration on the mobilisation of Mn during the filtration process. It can be observed that fulvic compounds have the highest capacity of releasing Mn into the filtrate water. The concentration of Mn increased from $250\pm47 \mu g/L$ to $1041\pm69 \mu g/L$ as the concentration of fulvic acid increased from 5 mg-C/L to 20 mg-C/L. The same trend was noted for humic compounds, where a humic acid concentration of 20 mg-



Figure 4. Effect of NOM type (humic, fulvic, and tyrosine) and concentration (5, 10, 15, 20 mg-C/L) on (a) Fe, (b) Mn, and (c) As mobilisation during the batch experiment (anaerobic, 30 °C, HRL = 0.5 m/d)

C/L resulted in increasing the released Mn concentration five times compared with its effluent concentration in the reactors injected with 5 mg-C/L humic acid. Likewise, a positive relationship was observed between Mn concentration in the batch reactor effluents and tyrosine concentration. The concentration of Mn was increased by 35%, 65%, and 77% in the batch reactors injected with 10, 15, and 20 mg-C/L, respectively, compared with its concentration in
the batch reactors injected with 5 mg-C/L.

The batch study results showed that the mobilisation of As is obviously affected by the type 386 and concentration of NOM (Figure 4.C). NOM at low concentration exhibited a poor capacity 387 of releasing As from its precipitated form into the filtrate water. No As concentration was 388 389 quantitatively (LOQ = $2 \mu g/L$) detected in any of the batch reactor effluents at DOC = 5 mg-C/L for the three NOM types. At higher concentrations (20 mg-C/L), tyrosine exhibited the 390 highest capacity of releasing As into the filtrate water; the As concentrations in the batch 391 reactors injected with humic, fulvic, and tyrosine were 3.6±0.4, 4.1±0.3, and 5.4±0.6 µg/L, 392 respectively. 393

394 4. Discussion

395 4.1. Mobilisation of Fe, Mn and As during BF

The mobilisation of Mn was two orders of magnitude higher than that of Fe during the infiltration process under the experimental conditions. From stoichiometry, 1 mole of simple organic matter (e.g., acetate) could act as an electron donor to mobilise 4 moles of Mn and 8 moles of Fe from the soil into the filtrate water (Lovley et al., 1988). However, the higher mobilisation of Mn compared with those of Fe and As observed during this study was probably due the combination of three factors:

402 (i) The reducing environment was still not high enough to be dominated by the Fe microbial403 reduction process.

(ii) Mn oxide was used as an agent for the Fe oxidation process and thereby could be
precipitated as Fe oxides. This process would increase the re-adsorption of As on the surface
of iron oxides and thus reduce the concentrations of Fe and As in the filtrate water (Vega et al.,

2017). Neidhardt et al. (2014) illustrated that Fe (oxyhydr)oxide redox processes control theconcentration of As in filtrate water.

(iii) The column experiment was conducted at a high temperature (30 °C) and a low infiltration
rate (0.5 m/d), which greatly enhanced the microbial reduction of manganese.

411 Low infiltration rate increases the contact time and interaction between the metals of the soil and the DOM of the raw water. Moreover, it promotes the reducing conditions along the flow 412 path, which leads to increase the mobilisation of metals into the infiltrating water. Paufler et 413 al. (2018) conducted column studies to assess the impact of the temperature (15–35 °C) and 414 flow rate on the Mn mobilisation during the BF process. It was found that raising the 415 temperature from 20 to 30 °C at a low hydraulic rate (1 ml/min) resulted in increasing the Mn 416 417 release rate (K_{Mn}) 10–15 times, whereas K_{Mn} was negligible when the feed-water temperature 418 was below 10 °C. Bourg et al. (1994) reported that increasing the temperature (>10 °C) triggers the microbial reduction of manganese in an alluvial aquifer. Based on these findings, it is 419 420 presumed that the higher Mn mobilisation rate in the column experiment relative to Fe and As 421 mobilisation is primarily ascribable to the microbial reduction of Mn at high temperature (30 °C) that stimulates simultaneously the associated Fe (II) and As (III) oxidation and precipitation 422 processes. This plausible mechanism was observed and reported in many BF and alluvial 423 aquifer fields (Bourg et al., 1993, 1994; Eckert et al., 2006; Jacobs et al., 1988; Kedziorek et 424 al., 2008; Thomas et al., 1994; Vega et al., 2017). However, it is worth noting that other 425 mechanisms (e.g., the simultaneous mobilisation of Fe and Mn) have also been observed in 426 many BF fields (Grischek et al., 2017; Hamdan et al., 2013; Matsunaga et al., 1993). 427

428 4.2. Influence of composition and concentration of DOM on metal(loid) mobilisation

429 The strong correlation between DOC and metal(loid) (Fe, Mn, and As) concentrations in the 430 effluent water of the column indicates that DOM plays a key role in the mobilisation of these 431 metal(loid)s during infiltration. In this research, NCTW with low organic content demonstrated lower efficiency in releasing Fe, Mn, and As into the filtrate water compared with those of 432 other source-water types. This is consistent with the findings of recent studies (Hossain Md 433 434 Anawar et al., 2013; Vega et al., 2017), which reported an increase in the mobilisation rate of Fe and As associated with higher DOM in Daudkandi (Meghna delta) and Marua of the Ganges 435 delta plain aquifer in Bangladesh (Hossain M. Anawar et al., 2003; Hossain Md Anawar et al., 436 2013). Wang et al. (2012) illustrated that DOM influences the mobilisation and release of 437 metal(loid)s during the passage of the soil through a redox process, complexation, and 438 439 competitive adsorption. Therefore, it can be concluded that the organic composition of feed water controls the concentration of Fe, Mn, and As in bank filtrate water. 440

Refractory compounds (i.e., humic) play a vital role in the mobilisation of Fe, Mn, and As 441 442 during the subsurface flow of water. The experimental results revealed that the mobilisation of Fe and Mn has a strong positive relationship with the humic content of the feed water (HIX), 443 whereas As mobilisation was the least affected by changes in the humic content of the feed 444 water. Several previous studies highlighted the multiple roles of humic compounds in the 445 mobilisation of metal(loid)s during the filtration process (Brune et al., 2004; Liu et al., 2011; 446 447 Yuan et al., 2018). Owing to their high electron shuttle capacities, humic compounds might act as catalysts for the iron microbial reduction process by transferring electrons between the 448 449 insoluble metal(loid)s and the reducing microorganisms; this was investigated in many field 450 studies (Mladenov et al., 2010; Mladenov et al., 2015; Poggenburg et al., 2018). In addition, these electron-rich compounds have high capabilities of binding to the soil metal(loid)s and 451 increasing their solubility (Sharma et al., 2010). Chen et al. (2003) pointed out that humic 452 453 compounds are more soluble at higher pH (>4); thus, they are subjected to more trapping of metal(loid)s from the soil and are maintained in their soluble forms by forming soluble complex 454 compounds. 455

456 The effect of humic compounds on the metal(loid) mobilisation is relatively dependent on the origin and characteristics of the organic compounds and type of metal(loid). In this research, 457 a moderate correlation was detected between RI and Fe concentration in the filtrate water, 458 459 indicating that terrestrial humic compounds (with condensed structures) have relatively higher capability to trap Fe into filtrate water than lower molecular weight humic compounds. 460 According to Yuan et al. (2018), high-molecular-weight humic compounds have more binding 461 sites, and therefore higher capability of forming chemically stable compounds with Fe, than 462 less-condensed-structure and labile compounds. Therefore, it can be deduced that humic 463 464 compounds of terrestrial origin are more triggered to release iron from its solid forms into filtrate water. This was confirmed by the batch experiment results, which showed a relatively 465 higher increase in Fe mobilisation with humic than with fulvic acid (lower-molecular-weight 466 467 humic compounds). By contrast, a weak correlation was observed between RI and Mn mobilisation, demonstrating comparable abilities of humic compounds with low and high 468 condensation (i.e., terrestrial and microbial humic) to mobilise Mn; this indicates that Mn metal 469 470 has a high affinity to both low- and high-molecular-weight humic compounds. This is in agreement with Vega et al. (2017), who observed higher concentrations of Mn to be associated 471 with higher humic content in shallow aquifers at West Bengal (India). Nevertheless, the batch 472 experiment showed that fulvic compounds at higher concentrations (>5 mg-C/L) have a higher 473 474 capability of mobilising Mn than higher-molecular-weight humic compounds; this could be 475 due to their high capacity to form soluble and stable Mn complexes at high pH (>6) compared with humic compounds, as proven previously by Du Laing (2010). 476

The mobilisation of Mn during the filtration process was observed to be highly influenced by the biodegradable organic matter content of the source water. The high correlation between Mn concentration in the filtrate water and protein-like fluorescent components indicates that the microbial reduction of Mn is the primary mechanism of Mn release. However, a moderate 481 correlation was observed between Fe concentration and protein-like components. The batch experiment showed that a protein-like (i.e., tyrosine) compound is the most effective NOM for 482 releasing Fe and As at high concentrations (>10 mg-C/L). This is probably due to the increased 483 484 concentration of biodegradable compounds, which increases the biological activity associated to sand, increases the mobilisation of Mn and decreases the redox potential to a level that 485 permits bacteria to use Fe from the soil as a source of energy, thus increasing the Fe 486 487 concentration in the effluent water. This process is concurrent with the release of As adsorbed on Fe (oxyhydr)oxides into the filtrate water. 488

489 5. Conclusions

The mobilisation of geogenic metal(loid)s, such as Fe, Mn, and As, during the BF process restricts their application, particularly in arid-climate countries. Based on the results of laboratory-scale column and batch studies, which were conducted to assess the impact of the organic composition of raw water on the mobilisation of these metal(loids) during anaerobic BF, the following conclusions are drawn:

- The mobilisation of Mn under the applied experimental conditions was two and three
 orders of magnitude higher than those of Fe and As, respectively.
- 497 The dissolved organic matter concentration and composition of the raw water could
 498 change the capacity of mobilisation of metal(loids) during the filtration process
- The humic content of the source water was found to significantly affect the release of
 Fe and Mn during the BF process; a positive relationship was found between the
 mobilisation of the metal(loids) and the HIX of the source water. This might be
 attributed to the high electron shuttle capacity of the humic compounds, which enables
 them to act as mediators for the microbial reduction processes and to form soluble
 complex compounds with the released metals.

Terrestrial humic compounds with complex structures showed a higher capability of releasing Fe from the soil into the filtrate water. On the other hand, source water with lower-molecular-weight humic compounds at a concentration of >5 mg-C/L was able to mobilise Mn at a higher rate than higher-molecular-weight humic compounds.
 However, the metal(loid) species in the soil may alter the behaviours of NOM compounds, which should be investigated.

Arsenic mobilisation was observed to have a high correlation with Fe mobilisation and
 a weak correlation with the variations in the organic composition of source water.

Biodegradable organic matter was found to be effective in mobilising Mn into the filtrate water; a strong correlation was observed between the FI of protein-like components in the source water and Mn mobilisation. However, a moderate correlation was found between Fe mobilisation and the FI of protein-like components. Nevertheless, the experimental results showed that a high concentration (>10 mg-C/L) of a protein-like compound is enough to produce a Fe-reducing environment in the infiltration area and thereby increase the Fe concentration in the filtrate water.

In summary, this study revealed that the organic matter composition of source water
 determines the redox environment during the BF process and affects the mobilisation
 process of the metal(loid)s that should be considered during the BF design process.
 Moreover, this research highlights the efficiency of the fluorescence spectroscopy
 technique as a monitoring tool for characterisation the organic composition of the raw
 water as well as for prediction and control the redox process and the mobilisation of
 metal(loid)s during BF.

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- 531 6. References
- Abdelrady, A., Sharma, S., Sefelnasr, A., Abogbal, A., & Kennedy, M. (2019). Investigating the impact
 of temperature and organic matter on the removal of selected organic micropollutants during
 bank filtration: A batch study. *Journal of Environmental Chemical Engineering*, 7(1), 102904.
 doi:<u>https://doi.org/10.1016/j.jece.2019.102904</u>
- Abdelrady, A., Sharma, S., Sefelnasr, A., & Kennedy, M. (2018). The Fate of Dissolved Organic Matter
 (DOM) During Bank Filtration under Different Environmental Conditions: Batch and Column
 Studies. *Water*, 10(12), 1730.
- Anawar, H. M., Akai, J., Komaki, K., Terao, H., Yoshioka, T., Ishizuka, T., ... Kato, K. (2003). Geochemical occurrence of arsenic in groundwater of Bangladesh: sources and mobilization processes.
 Journal of Geochemical Exploration, 77(2), 109-131. doi:<u>https://doi.org/10.1016/S0375-6742(02)00273-X</u>
- Anawar, H. M., Tareq, S. M., & Ahmed, G. (2013). Is organic matter a source or redox driver or both
 for arsenic release in groundwater? *Physics and Chemistry of the Earth, Parts A/B/C, 58-60*,
 49-56. doi:https://doi.org/10.1016/j.pce.2013.04.009
- Baghoth, S. A., Sharma, S. K., & Amy, G. L. (2011). Tracking natural organic matter (NOM) in a drinking
 water treatment plant using fluorescence excitation-emission matrices and PARAFAC. *Water Res, 45*(2), 797-809. doi:10.1016/j.watres.2010.09.005
- Bertelkamp, C., Reungoat, J., Cornelissen, E., Singhal, N., Reynisson, J., Cabo, A., . . . Verliefde, A. R.
 (2014). Sorption and biodegradation of organic micropollutants during river bank filtration: A
 laboratory column study. *Water Research*, 52, 231-241.
 doi:http://dx.doi.org/10.1016/j.watres.2013.10.068
- 553Bourg, A. C. M., & Bertin, C. (1993). Biogeochemical processes during the infiltration of river water554into an alluvial aquifer. Environmental science & technology, 27(4), 661-666.555doi:10.1021/es00041a009
- Bourg, A. C. M., & Bertin, C. (1994). Seasonal and Spatial Trends in Manganese Solubility in an Alluvial
 Aquifer. *Environmental science & technology*, 28(5), 868-876. doi:10.1021/es00054a018
- Brune, A., Schink, B., Benz, M., & Kappler, A. (2004). Electron shuttling via humic acids in microbial
 iron(III) reduction in a freshwater sediment. *FEMS Microbiology Ecology*, 47(1), 85-92.
 doi:10.1016/S0168-6496(03)00245-9 %J FEMS Microbiology Ecology
- 561 Chen, J., Gu, B., Royer, R. A., & Burgos, W. D. (2003). The roles of natural organic matter in chemical
 562 and microbial reduction of ferric iron. *Science of The Total Environment, 307*(1), 167-178.
 563 doi:<u>https://doi.org/10.1016/S0048-9697(02)00538-7</u>
- 564Coble, P. G. (1996). Characterization of marine and terrestrial DOM in seawater using excitation-565emission matrix spectroscopy. Marine Chemistry, 51(4), 325-346.566doi:https://doi.org/10.1016/0304-4203(95)00062-3
- Du Laing, G. (2010). Analysis and Fractionation of Trace Elements in Soils. In P. S. Hooda (Ed.), *Trace Elements in Soils*.
- Eckert, P., & Irmscher, R. (2006). Over 130 years of experience with Riverbank Filtration in Düsseldorf,
 Germany. Journal of Water Supply: Research and Technology AQUA, 55(4), 283-291.
 doi:10.2166/aqua.2006.040
- Gabor, R. S., Baker, A., McKnight, D. M., & Miller, M. P. (2014). Fluorescence Indices and Their
 Interpretation. In A. Baker, D. M. Reynolds, J. Lead, P. G. Coble, & R. G. M. Spencer (Eds.),
 Aquatic Organic Matter Fluorescence (pp. 303-338). Cambridge: Cambridge University Press.
- 575 Ghodeif, K., Grischek, T., Bartak, R., Wahaab, R., & Herlitzius, J. (2016). Potential of river bank filtration 576 (RBF) in Egypt. *Environmental Earth Sciences*, *75*(8), 1-13. doi:10.1007/s12665-016-5454-3

- Gonçalves-Araujo, R., Granskog, M. A., Bracher, A., Azetsu-Scott, K., Dodd, P. A., & Stedmon, C. A.
 (2016). Using fluorescent dissolved organic matter to trace and distinguish the origin of Arctic
 surface waters. *Scientific Reports, 6*, 33978. doi:10.1038/srep33978
- 580 Grischek, T., & Paufler, S. (2017). Prediction of Iron Release during Riverbank Filtration. *9*(5), 317.
- Haas, R., Opitz, R., Grischek, T., & Otter, P. (2018). The AquaNES Project: Coupling Riverbank Filtration
 and Ultrafiltration in Drinking Water Treatment. *Water*, *11*(1), 18.
- Hamdan, A. M., Sensoy, M. M., & Mansour, M. S. (2013). Evaluating the effectiveness of bank
 infiltration process in new Aswan City, Egypt. *Arabian Journal of Geosciences, 6*(11), 41554165. doi:10.1007/s12517-012-0682-7
- Hansen, A. M., Kraus, T. E. C., Pellerin, B. A., Fleck, J. A., Downing, B. D., & Bergamaschi, B. A. (2016).
 Optical properties of dissolved organic matter (DOM): Effects of biological and photolytic degradation. *61*(3), 1015-1032. doi:doi:10.1002/lno.10270
- Harshman, R. A., & Lundy, M. E. (1994). PARAFAC: Parallel factor analysis. *Computational Statistics & Data Analysis*, 18(1), 39-72. doi:<u>https://doi.org/10.1016/0167-9473(94)90132-5</u>
- 591 Hinkle, D. E., Wiersma, W., & Jurs, S. G. (1988). Applied statistics for the behavioral sciences.
- Huguet, A., Vacher, L., Relexans, S., Saubusse, S., Froidefond, J. M., & Parlanti, E. (2009). Properties of
 fluorescent dissolved organic matter in the Gironde Estuary. *Organic Geochemistry*, 40(6),
 706-719. doi:<u>https://doi.org/10.1016/j.orggeochem.2009.03.002</u>
- Irmscher, R., & Teermann, I. (2002). Riverbank filtration for drinking water supply a proven method,
 perfect to face today's challenges. *Water Science and Technology: Water Supply, 2*(5-6), 1-8.
 doi:10.2166/ws.2002.0143 %J Water Science and Technology: Water Supply
- 598Jacobs, L. A., von Gunten, H. R., Keil, R., & Kuslys, M. (1988). Geochemical changes along a river-599groundwater infiltration flow path: Glattfelden, Switzerland. Geochimica et Cosmochimica600Acta, 52(11), 2693-2706. doi:https://doi.org/10.1016/0016-7037(88)90038-5
- Kedziorek, M. A., Geoffriau, S., Bourg, A. C. J. E. s., & technology. (2008). Organic matter and modeling
 redox reactions during river bank filtration in an alluvial aquifer of the Lot River, France. 42(8),
 2793-2798.
- Li, P., Lee, S. H., Lee, S. H., Lee, J.-B., Lee, Y. K., Shin, H.-S., & Hur, J. (2016). Seasonal and storm-driven
 changes in chemical composition of dissolved organic matter: a case study of a reservoir and
 its forested tributaries. *Environmental Science and Pollution Research*, 23(24), 24834-24845.
 doi:10.1007/s11356-016-7720-z
- Liu, G., Fernandez, A., & Cai, Y. (2011). Complexation of Arsenite with Humic Acid in the Presence of Ferric Iron. *Environmental science & technology, 45*(8), 3210-3216. doi:10.1021/es102931p
- Lovley, D. R., & Phillips, E. J. (1988). Novel mode of microbial energy metabolism: organic carbon
 oxidation coupled to dissimilatory reduction of iron or manganese. *Applied and Environmental Microbiology*, 54(6), 1472-1480.
- Massmann, G., Nogeitzig, A., Taute, T., & Pekdeger, A. (2008). Seasonal and spatial distribution of
 redox zones during lake bank filtration in Berlin, Germany. *Environmental Geology*, 54(1), 53 65. doi:10.1007/s00254-007-0792-9
- Matsunaga, T., Karametaxas, G., Von Gunten, H., & Lichtner, P. J. G. e. C. A. (1993). Redox chemistry
 of iron and manganese minerals in river-recharged aquifers: A model interpretation of a
 column experiment. 57(8), 1691-1704.
- Miller, M. P., McKnight, D. M., Cory, R. M., Williams, M. W., & Runkel, R. L. (2006). Hyporheic Exchange
 and Fulvic Acid Redox Reactions in an Alpine Stream/Wetland Ecosystem, Colorado Front
 Range. *Environmental science & technology*, 40(19), 5943-5949. doi:10.1021/es060635j
- Mladenov, N., Zheng, Y., Miller, M. P., Nemergut, D. R., Legg, T., Simone, B., ... McKnight, D. M. (2010).
 Dissolved Organic Matter Sources and Consequences for Iron and Arsenic Mobilization in
 Bangladesh Aquifers. *Environmental science & technology*, 44(1), 123-128.
 doi:10.1021/es901472g
- Mladenov, N., Zheng, Y., Simone, B., Bilinski, T. M., McKnight, D. M., Nemergut, D., . . . Ahmed, K. M.
 (2015). Dissolved Organic Matter Quality in a Shallow Aquifer of Bangladesh: Implications for

- 628ArsenicMobility.Environmentalscience& technology,49(18),10815-10824.629doi:10.1021/acs.est.5b01962
- Murphy, K. R., Stedmon, C. A., Graeber, D., & Bro, R. (2013). Fluorescence spectroscopy and multi-way
 techniques. PARAFAC. *Analytical Methods*, 5(23), 6557-6566. doi:10.1039/C3AY41160E
- Neidhardt, H., Berner, Z. A., Freikowski, D., Biswas, A., Majumder, S., Winter, J., ... Norra, S. (2014).
 Organic carbon induced mobilization of iron and manganese in a West Bengal aquifer and the
 muted response of groundwater arsenic concentrations. *Chemical Geology*, *367*, 51-62.
 doi:https://doi.org/10.1016/j.chemgeo.2013.12.021
- Ohno, T. (2002). Fluorescence Inner-Filtering Correction for Determining the Humification Index of
 Dissolved Organic Matter. *Environmental science & technology, 36*(4), 742-746.
 doi:10.1021/es0155276
- Paufler, S., Grischek, T., Benso, M. R., Seidel, N., & Fischer, T. (2018). The Impact of River Discharge
 and Water Temperature on Manganese Release from the Riverbed during Riverbank
 Filtration: A Case Study from Dresden, Germany. *10*(10), 1476.
- Poggenburg, C., Mikutta, R., Schippers, A., Dohrmann, R., & Guggenberger, G. (2018). Impact of natural
 organic matter coatings on the microbial reduction of iron oxides. *Geochimica et Cosmochimica Acta, 224*, 223-248. doi:https://doi.org/10.1016/j.gca.2018.01.004
- Qian, C., Wang, L.-F., Chen, W., Wang, Y.-S., Liu, X.-Y., Jiang, H., & Yu, H.-Q. (2017). Fluorescence
 Approach for the Determination of Fluorescent Dissolved Organic Matter. *Analytical Chemistry*, 89(7), 4264-4271. doi:10.1021/acs.analchem.7b00324
- R.K. Pachauri, & Meyer, L. A. (2014). *Climate Change 2014 Synthesis Report*. Retrieved from Geneva,
 Switzerland:
- Ray, C., & Shamrukh, M. (2011). Riverbank Filtration for Water Security in Desert Countries. *NATO Science for Peace and Security Series C: Environmental Security, 2011.*
- Sandhu, C., Grischek, T., Börnick, H., Feller, J., & Sharma, S. K. (2019). A Water Quality Appraisal of
 Some Existing and Potential Riverbank Filtration Sites in India. *11*(2), 215.
- Schittich, A.-R., Wünsch, U. J., Kulkarni, H. V., Battistel, M., Bregnhøj, H., Stedmon, C. A., & McKnight,
 U. S. (2018). Investigating Fluorescent Organic-Matter Composition as a Key Predictor for
 Arsenic Mobility in Groundwater Aquifers. *Environmental science & technology*, *52*(22),
 13027-13036. doi:10.1021/acs.est.8b04070
- Sharma, P., Ofner, J., & Kappler, A. (2010). Formation of Binary and Ternary Colloids and Dissolved
 Complexes of Organic Matter, Fe and As. *Environmental science & technology*, 44(12), 4479 4485. doi:10.1021/es100066s
- Singh, S., Inamdar, S., & Scott, D. (2013). Comparison of Two PARAFAC Models of Dissolved Organic
 Matter Fluorescence for a Mid-Atlantic Forested Watershed in the USA. *Journal of Ecosystems*,
 2013, 16. doi:10.1155/2013/532424
- Sprenger, C., Lorenzen, G., & Asaf, P. (2012). Environmental Tracer Application and Purification
 Capacity at a Riverbank Filtration Well in Delhi (India). *Journal of Indian Water Works* Association, Special Issue on River Bank Filtration, 1, 25-32.
- Thomas, N. E., Kan, K. T., Bray, D. I., & MacQuarrie, K. T. B. (1994). Temporal Changes in Manganèse
 Concentrations in Water from the Fredericton Aquifer, New Brunswick. 32(4), 650-656.
 doi:doi:10.1111/j.1745-6584.1994.tb00901.x
- Vega, M. A., Kulkarni, H. V., Mladenov, N., Johannesson, K., Hettiarachchi, G. M., Bhattacharya, P., . . .
 Datta, S. (2017). Biogeochemical Controls on the Release and Accumulation of Mn and As in
 Shallow Aquifers, West Bengal, India. *Frontiers in Environmental Science*, *5*, 29.
- Wahaab, R. A., Salah, A., & Grischek, T. (2019). Water Quality Changes during the Initial Operating
 Phase of Riverbank Filtration Sites in Upper Egypt. *Water, 11*(6), 1258.
- Wang, Y., Jiao, J. J., & Cherry, J. A. (2012). Occurrence and geochemical behavior of arsenic in a coastal
 aquifer–aquitard system of the Pearl River Delta, China. *Science of The Total Environment*,
 427-428, 286-297. doi:<u>https://doi.org/10.1016/j.scitotenv.2012.04.006</u>

- Wünsch, U., Murphy, K., & Stedmon, C. (2017). The One-Sample PARAFAC Approach Reveals
 Molecular Size Distributions of Fluorescent Components in Dissolved Organic Matter.
 Environmental Science and Technology, *51*(20), 11900–11908. doi:10.1021/acs.est.7b03260
- Yang, C., Li, S., Liu, R., Sun, P., & Liu, K. (2015). Effect of reductive dissolution of iron (hydr)oxides on
 arsenic behavior in a water-sediment system: First release, then adsorption. *Ecological Engineering*, 83, 176-183. doi:10.1016/j.ecoleng.2015.06.018
- Yuan, Y., He, X., Xi, B., Li, D., Gao, R., Tan, W., . . . Zhao, X. (2018). Polarity and molecular weight of
 compost-derived humic acid affect Fe(III) oxides reduction. *Chemosphere, 208*, 77-83.
 doi:<u>https://doi.org/10.1016/j.chemosphere.2018.05.160</u>