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Corrigendum

Corrigendum to “*Natronobiforma cellulositropha* gen. nov., sp. nov., a novel haloalkaliphilic member of the family *Natrialbaceae* (class *Halobacteria*) from hypersaline alkaline lakes” [Syst. Appl. Microbiol. 41 (2018) 355–362]

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This is a corrigendum to the protologue **Table 3** describing properties of *Natronobiforma cellulositropha* gen. nov sp. nov.

Table 3
Natronobiforma cellulositropha: protologue.

Parameter	Genus: <i>Natronobiforma</i> gen. nov.	Species: <i>Natronobiforma cellulositropha</i> sp. nov.
Date created	2018-03-04	2018-03-04
Taxon number (TXNR)	TA00433	TA00433
Author (AUTE)	Dimitry Y. Sorokin	
Species name (SPNA)	<i>Natronobiforma</i>	<i>Natronobiforma cellulositropha</i>
Genus name (GENA)	—	<i>cellulositropha</i>
Specific epithet (SPEP)	—	sp. nov.
Species status (SPST)	<i>Natronobiforma</i> (Na.tro.no.bi.for'ma Gr. neut. n. <i>natron</i> , arbitrarily derived from the Arabic n. <i>natrun</i> or <i>natron</i> , soda; L. adv. num. <i>bis</i> , twice; L. fem. n. <i>forma</i> , form, shape; N.L. fem. n. <i>Natronobiforma</i> , the dimorphic natronoarchaeon	(cel.lu.lo.si.tro'pha N.L. n. <i>cellulosum</i> , cellulose; N.L. fem. n. <i>trophē</i> nourishment, food; N.L. fem. adj. <i>cellulositropha</i> , utilizer of cellulose)
Etymology (GETY/SPTY)	Dimitry Y. Sorokin, Tatiana V. Khijniak, Nadezhda A. Kostrikina, Alexander G. Elcheninov, Stepan V. Toshchakov, Nicole J. Bale, Jaap S. Sinninghe Damsté, Ilya V. Kublanov	
Authors (AUT)	<i>Natronobiforma cellulositropha</i> gen. nov., sp. nov., a novel haloalkaliphilic member of the family <i>Natrialbaceae</i> (class <i>Halobacteria</i>) from hypersaline alkaline lakes	
Title (TITL)	Systematic and Applied Microbiology	
Journal (JOUR)	Dimitry Y. Sorokin	
Corresponding author (COAU)	d.sorokin@tudelft.nl ; soroc@immi.ru	
E-mail of corresponding author (EMAU)		AArcel5
Designation of the type strain (TYPE)	—	JCM 31939; UNIQEM U972
Strain collection numbers (COLN)	—	KT247980
16S rRNA gene accession number (16 SR)	—	<i>rpoB</i> ' [MG940906]
Alternative house-keeping genes: gene [accession numbers] (HKGN)	—	

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Table 3 (Continued)

Genome status (GSTA)	–	draft
GC mol% (GGCM)	–	65.4–65.5 (genomes of AArcel5 ^T and AArcel2)
Country of origin (COUN)	Russian Federation	
Region of origin (REGI)	Altai region	
Date of isolation (DATI)	2013-08-15	
Source of isolation (SOUR)	Surface sediments and brines of hypersaline alkaline lakes	Surface sediments from hypersaline soda lake Tanatar-1
Sampling dates (DATS)	2013-07-07	
Geographic location (GEOL)	S-W Siberia, N-E Mongolia, California	
Latitude (LATI)	51°39'N	51°39'N
Longitude (LONG)	79°48'E	79°48'E
Depth (DEPT)	0.1 m	0.1 m
Temperature of the sample (TEMS)	25 °C	25 °C
pH of the sample (PHSA)	11.0	11.0
Salinity of the sample (SALS)	40%	40%
Number of strains in study (NSTR)	6	
Source of isolation of non-type strains (SAMP)	Hypersaline alkaline lakes in Russia, Mongolia and California	
Growth medium, incubation conditions (CULT)	Alkaline medim containing 4 M Na ⁺ with pH 9–9.5 and cellulose as substrate	4 M total Na ⁺ , equal mix of sodium carbonate and NaCl on the basis of Na molarity, pH 9.5; incubation – 37 °C; amorphous cellulose or cellobiose as C and energy source
Conditions of preservation (PRES)	Deep freezing in 15% glycerol (v/v)	
Gram stain (GRAM)	Negative	
Cell shape (CSHA)	Pleomorphic, from flat motile rods to nonmotile coccoid cells	
Cell size (CSZI)	–	0.5–0.8 µm in diameter, length is variable
Motility (MOTY)	–	Motile
Motility type (MOTK)	–	Flagellar
Type of flagellation (TFLA)	–	Variable, from single subpolar to several peritrichous flagella
Sporulation (SPOR)	None	
Colony morphology (COLM)	–	Pink, up to 2 mm
Temperature range for growth (TEMR)	–	20–53 °C
Lowest temperature for growth (TEML)	–	20
Highest temperature for growth(TEMH)	–	53
Optimal temperature for growth (TEMO)	–	43
Lowest pH for growth (PHLO)	–	7.5
Highest pH for growth (PHHI)	–	9.9
Optimum pH for growth (PHOP)	–	9.0
pH category (PHCA)	Alkaliphile (optimum >8.5)	
Lowest NaCl concentration for growth (SALL)	–	2.5
Highest NaCl concentration for growth (SALH)	–	4.8
Optimum salt concentration for growth (SALO)	–	4.0
Other salts important for growth	Sodium carbonates	
Salinity category (SALC)	Extreme halophilic (optimum 4 M Na ⁺)	
Relation to oxygen (OREL)	Aerobe	
O ₂ conditions for strain testing (OCON)	Aerobic	
Carbon source used (class) (CSUC)	Carbohydrates	
Specific compounds (CSUC)	Cellulose, xylan, mannan, cellobiose, maltose	
Nitrogen source (NSOU)	Ammonium	
Terminal electron acceptor (ELAC)	O ₂	
Energy metabolism (EMET)	Chemoorganotrophic	
Phospholipids (PHOS)	Core membrane lipids are archaeol (C20–C20 DGE) and C20–C25 DGE in equal proportion	Phosphatidylglycerophosphate methyl ester (PGP-Me), phosphatidylglycerol (PG), phosphatidylglycerol sulfate (PGS) and phosphatidylglycerophosphate (PGP) Phosphatidylglycose (GL-PG), diglycosyl (2GL)
Glycolipids (GLYC)	–	
Habitat (HABT)	Hypersaline alkaline lakes	
Extraordinary features (EXTR)	Growth with native insoluble cellulose	Fast growth with insoluble native celluloses; more than 30 GH glucosyl-hydrolases genes in the genome

(-) not specific for the genus.

1. The species name “cellulotropha” was corrected to “cellulositropha” in the (SPNA), (SPEP) and TITL lines.
2. The author name “Damst  d” in the (AUT) was corrected to “Damst  ”.
3. The word “neutral” in the (GETY/SPTY) was corrected to “neut”.
4. Some of the relevant values have been added to both genus and species columns.