

## **The Role of Insects in Medical Engineering and Bionics Towards Entomomedical Engineering**

Bloemberg, Jette; Stefanini, Cesare; Romano, Donato

**DOI**

[10.1109/TMRB.2021.3101693](https://doi.org/10.1109/TMRB.2021.3101693)

**Publication date**

2021

**Document Version**

Final published version

**Published in**

IEEE Transactions on Medical Robotics and Bionics

**Citation (APA)**

Bloemberg, J., Stefanini, C., & Romano, D. (2021). The Role of Insects in Medical Engineering and Bionics: Towards Entomomedical Engineering. *IEEE Transactions on Medical Robotics and Bionics*, 3(4), 909-918. <https://doi.org/10.1109/TMRB.2021.3101693>

**Important note**

To cite this publication, please use the final published version (if applicable). Please check the document version above.

**Copyright**

Other than for strictly personal use, it is not permitted to download, forward or distribute the text or part of it, without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license such as Creative Commons.

**Takedown policy**

Please contact us and provide details if you believe this document breaches copyrights. We will remove access to the work immediately and investigate your claim.

***Green Open Access added to TU Delft Institutional Repository***

***'You share, we take care!' - Taverne project***

**<https://www.openaccess.nl/en/you-share-we-take-care>**

Otherwise as indicated in the copyright section: the publisher is the copyright holder of this work and the author uses the Dutch legislation to make this work public.

# The Role of Insects in Medical Engineering and Bionics: Towards Entomomedical Engineering

Jette Bloemberg<sup>1b</sup>, Cesare Stefanini, *Member, IEEE*, and Donato Romano<sup>1b</sup>

**Abstract**—Insects are important agents in ecosystems. Their diverseness and developed coping mechanisms also make them interesting for direct application and as a source of inspiration in medical engineering. We summarized the main contribution of insects in biomedical applications. Medical centers in North America, and Europe use fly larvae for maggot therapy to remove necrotic tissue, decrease infection risk, and improve wound healing. Ant mandibles are used as a suturing technique by African tribes and as sources of inspiration for surgical clamps. Both the mosquito fascicle and the wasp ovipositor are sources of inspiration for the design of medical needles. Herein, a new research field called “*entomomedical engineering*,” is proposed. We define entomomedical engineering as the branch of engineering that uses insects either directly or as a source of inspiration to design and develop medical treatments or instruments. In addition, we want to emphasize the importance of preserving insects because of their function in the ecosystem, medicine, and medical engineering.

**Index Terms**—Biologically-inspired design, insect, maggot therapy, medical clamp, medical needle.

## I. INTRODUCTION

**H**UMANS see many insect species as pests, but these organisms play a crucial role in keeping the planet livable [1], [2]. Insects are important providers [3], decomposers [4], pest controllers [5], pollinators [6], soil engineers [7], and more. Insects are a source of food for larger animals including other arthropods, fish, amphibians, reptiles, birds, and mammals. Without insects, species higher up in the food chain suffer from food scarcity [8]. Besides their essential roles in the ecosystem, clinicians and engineers use insects directly and as a source of inspiration [9], [10]. Insects are the most diverse organisms. They are unmatched in their species

Manuscript received May 19, 2021; revised June 18, 2021; accepted July 27, 2021. Date of publication August 2, 2021; date of current version November 19, 2021. This paper was recommended for publication by Associate Editor A. Ijspeert and Editor M. Mitsuishi upon evaluation of the reviewers’ comments. The work of Jette Bloemberg was supported by EU funds for the Erasmus+ Traineeship at host institution Sant’Anna School of Advanced Studies. (*Corresponding author: Donato Romano.*)

Jette Bloemberg is with the BioRobotics Institute, Sant’Anna School of Advanced Studies, 56025 Pisa, Italy, and also with the Faculty of Mechanical, Maritime, and Materials Engineering, Technical University of Delft, 2628 CD Delft, The Netherlands.

Cesare Stefanini is with the BioRobotics Institute and the Department of Excellence in Robotics and A.I., Sant’Anna School of Advanced Studies, 56025 Pisa, Italy, and also with the Healthcare Engineering Innovation Center, Khalifa University, Abu Dhabi, UAE.

Donato Romano is with the BioRobotics Institute and the Department of Excellence in Robotics and A.I., Sant’Anna School of Advanced Studies, 56025 Pisa, Italy (e-mail: donato.romano@santannapisa.it).

This article has supplementary downloadable material available at <https://doi.org/10.1109/TMRB.2021.3101693>, provided by the authors.

Digital Object Identifier 10.1109/TMRB.2021.3101693

numbers, biomass, and ecological impact [11]. Their cycle of reproduction resumes quickly, allowing the development of high genetic diversity. They developed characteristics and coping mechanisms to survive in extreme situations, making them attractive sources of direct application and bio-inspiration in medical engineering as bio-engineers. The direct application of insects or insect-derived products in medicine is called entomotherapy [12]–[14]. Insect-derived products such as honey, venom, and insect anticoagulants and their clinical application were described by Rather *et al.* [14]. Insects are used directly in bio-surgery and bio-clamps, which often already started in veterinary or traditional medicine and are recently becoming of increasing interest [15], [16].

Recent studies showed population declines of insect species [17], [18]. Other studies stated that terrestrial insects decline, whereas the number of freshwater insects increased [19]. Montgomery *et al.* [20] argue that most documented collapses of insect species are from geographically restricted studies, which do not allow us to conclude insect species’ declines on a global scale. There is a need for greater investigation of insect declines. However, the severity of the reported insect declines calls for immediate action [21]. The primary drivers of insect declines are pollution by pesticides and fertilizers, insect habitat loss and degradation, biological factors such as pathogens and introduced species, and climate change [22].

In this review, we summarize the main contribution of insects in biomedical applications. We included both the insects’ direct application in therapies as bio-actuators and the insects’ indirect application as a source of inspiration for biomedical tools. Furthermore, we want to emphasize the importance of preserving insects because of their ecological function, as well as of their role in medicine and medical engineering. This review provides an overview of the scientific literature on both the direct application and bio-inspiration of insects in medicine. The first part of this review describes the insects’ direct application in medicine, including bio-surgery with maggot debridement therapy (MDT) and bio-clamps. The second part describes the insect-inspired medical devices in the scientific literature: bio-inspired needles and bio-inspired clamps.

## II. MATERIALS AND METHODS

To survey the scientific literature of insects used in medicine, we used the Scopus database. The search query

was a Boolean combination of keywords regarding the following: (1) the type of insect and (2) the target application. In the Scopus search, we used the function “LIMIT TO” to limit the search to English language publications and publications within the “Medicine” and “Engineering” subject areas. We used a separate search query for bio-surgery, bio-clamps (including bio-inspired clamps), mosquito-inspired needles, and wasp-inspired needles. Supplementary file S1 contains the search queries used in this study.

The search resulted in 342 articles from the Scopus database. The title and abstract of the scientific articles were screened based on the eligibility criteria. For the articles about bio-surgery, solely articles describing controlled clinical trials of therapy using alive, disinfected fly larvae, i.e., maggots, were included. We excluded case studies and *in vitro* tests. For the articles on bio-clamps and bio-inspired needles, we included articles describing the mechanical working principle or design. We excluded articles focused on motion planning algorithms, computational modelling, or tissue-needle interaction. We read the full text of the remaining papers and checked the references of the articles included in this review to retrieve relevant articles not captured by the search query in Scopus. In the end, this review encompasses 93 scientific articles.

### III. DIRECT APPLICATION OF INSECTS IN MEDICINE

#### A. Bio-Surgery

Humans have used insects in skin wound healing for a long time, and our interest in their use has increased in the last years. The definition of bio-surgery is using living maggots to remove necrotic tissue, decrease the risk of infection, and improve wound healing [23], [24]. Other terms used in the scientific literature for bio-surgery are maggot therapy, larval therapy, maggot debridement therapy (MDT), larval debridement therapy, maggot wound therapy, and biodebridement [25]–[28]. Some fly species’ larvae feed upon living or decaying animal tissue called myiasis [29], [30]. The Diptera order is the main source of insects in direct bio-surgery applications. Some larvae limit their digestion to dead or necrotic tissue called semi-specific myiasis. These larvae are the ones that are used in bio-surgery. The “greenbottle” blowfly, *Lucilia sericata* Meigen (Diptera: Calliphoridae), is used the most in maggot therapy [31]. Other species often used are *Lucilia illustris* Meigen (Diptera: Calliphoridae) and *Phormia regina* Meigen (Diptera: Calliphoridae) [32]. The maggots produce proteolytic enzymes like collagenase [33], [34]. Collagenase effectively breaks down necrotic tissue to a semi-liquid, which the larvae absorb and digest.

Table I shows a timeline of the history of scientific literature on the use of maggot therapy. Ambroise Paré was the first to observe the beneficial effects of maggots in wounds during the battle of St. Quentin in 1557 [35]. Baer [36] was the first to report a scientific study on the application of maggots to treat chronic osteomyelitis [37]. During World War I, Baer, an orthopedic, found that the wounds of two soldiers swarmed with maggots. The wounds were granulated with

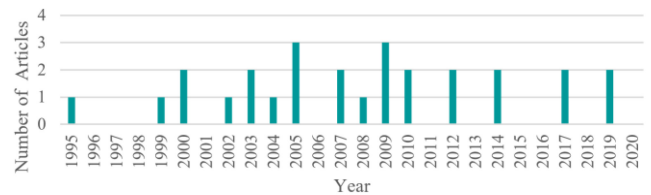


Fig. 1. Temporal distribution of articles about a clinical study that applies maggot therapy.

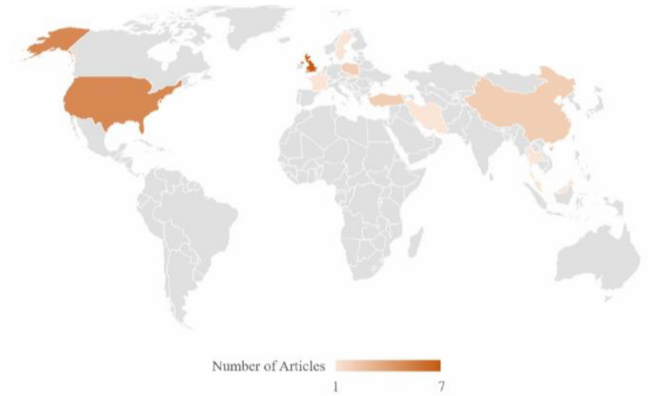


Fig. 2. Spatial distribution of the articles about a clinical study that applies maggot therapy.

no evidence of sepsis. MDT was popular in the 1930s and early 1940s to clean exposed wounds [30]. By the mid-1940s, maggot therapy ceased in popularity, simultaneously with the beginning of the antibiotic era [30], [38]. Antibiotics prevented the spread of infections by bacteria that could lead to soft-tissue complications, which the larvae treated previously [25]. MDT’s other main challenges were social approval, local pruritis, the use of sterile maggots, and wounds with copious exudate because then maggots die due to lack of oxygen. For more information about the history of maggot therapy, we refer the reader to [30], [32], [37]–[42].

At the end of the twentieth century, maggot therapy’s revival started with the first controlled clinical trials [43]. Also, the treatment of other wounds than osteomyelitis was studied. Clinicians extended MDT to the field of diabetes and land mine wounds in developing nations where access to medical treatment may be inadequate [38], [44]. Figures 1 and 2 show the temporal and spatial distribution of the articles published about a clinical study that applies maggot therapy, respectively. The advantages of maggots caused the revival, as they are (1) active in debriding necrotic tissue, (2) relative safe, (3) simple, (4) efficient, (5) cheap, and (6) effective even in the context of antibiotic-resistant infections [25], [38]. By 2000, MDT was a treatment option in approximately 50 medical centers in North America and the United Kingdom [38]. Furthermore, medical maggots were produced and distributed to 400 medical centers in the United Kingdom, Belgium, Germany, and Sweden [38]. In 2004, the U.S. Food and Drug Administration (FDA) granted market clearance to medicinal maggots as a medical device [45]. The maggot’s physical activity is essential for the debridement; therefore, the FDA classified the medical maggots as a medical device instead of a drug [43]. Maggots move over the wound,

TABLE I  
SCIENTIFIC LITERATURE ABOUT THE USE OF MAGGOTS IN MEDICINE UNTIL ITS POPULARITY CEASED IN 1940

Time	Event	Reference
1557 – Battle of St. Quentin	Abroise Paré observed the beneficial effects of fly larvae in wounds.	[35]
1929 – Syrian campaign	DJ Larray reported that maggots removed necrotic tissue, while enhancing the granulation of living tissue in the soldiers of Napoleon Bonaparte's army he treated.	[36]
1931 – World War I	Scientific study on the use of maggots for the treatment of chronic osteomyelitis. Method to rear sterile maggots, by first sterilizing the eggs and then raising the maggots on a sterile food source.	[36]
1934	Wound becomes alkaline, because the maggots excrete calcium carbonate.	[145]
1935	MDT is useful in any wound containing sloughing tissue which can be exposed by incision, not only for osteomyelitis. Maggot therapy is the most rapid method to remove sloughing soft tissue after excision.	[146]
1940s	MDT ceased in popularity, simultaneously with the beginning of the antibiotic era.	[30, 38]

MDT = maggot debridement therapy

thereby ploughing the tissue and simultaneously spreading its alimentary secretions and excretions (ASE). The ASE liquefies the necrotic tissue. Afterward, the maggots can imbibe it. Medicinal maggots work through the ASE and the physical contact with the host, inducing debridement, disinfection, growth stimulation, and wound healing. For more information about the maggot-host interactions to achieve these working mechanisms, we refer the reader to [43]. In the Netherlands, the larvae of *L. sericata* were approved as an unregistered medicine in 2014 [46]. In Germany, there were 5017 cases of MDT in 2016 [47]. Other countries where MDT is reintroduced are Israel, the United Kingdom, Sweden, Switzerland, Ukraine, and Thailand [47]. Supplementary file S2 presents clinical studies of wound treatment with maggots (*L. sericata*, unless otherwise specified). The studies in S2 show that the maggots' debridement efficacy is beyond doubt, yet the disinfection and growth stimulation activity remains questionable [43]. Sherman [43] stated we need more clarity regarding MDT's role in promoting wound closure. Maggots' physical effects do not last longer than a few weeks after MDT ended. If the wounds do not heal immediately, there is the risk for recolonization, infection, stagnation, and necrosis.

#### B. Bio-Clamp

Closing a wound using ant mandibles of large black ants is described as one of the first suturing techniques [48]–[54]. The insects family Formicidae of the order Hymenoptera is the main source of insects in direct bio-clamp applications. The clinician lets multiple ants bite the wound edges and pull them together [48], [49], [51]–[53]. Afterward, the ants' bodies are twisted off or cut off, leaving the head with the mandibles to maintain the edges close together [48], [49], [51]–[53]. Supplementary file S3 contains a timeline of the reported use of ants as a suturing technique. Schiappa and Van Hee [52] stated that ancient populations in various continents used giant ants like *Oecophylla smaragdina* (Fabricius) and *Eciton burchelli* (Westwood) as a suturing device. These ants have powerful claws that can draw the edges of a wound together by biting them. Iavazzo *et al.* [53] stated that different ant species were used in different parts of the world, amongst them *Atta cephalotes* (Linnaeus), *Eciton burchellii* (Westwood), and *Oecophylla smaragdina* (Fabricius). Nowadays, ant mandibles are used as a suturing technique by some African tribes [51].

#### IV. INSECTS AS INSPIRATION MODELS IN MEDICAL ENGINEERING

Mechanical failure in guidewires and needle-like instruments are often caused by buckling. To prevent buckling, either the penetration tool's critical load can be increased, or the substrate's penetration load can be decreased [55]. Sakes *et al.* [55] stated that the mosquito proboscis and wasp ovipositors are examples from nature that combine different buckling prevention strategies. The insect orders Diptera and Hymenoptera are the main sources of inspiration in bio-inspired needles.

##### A. Mosquito-Inspired Microneedles

Microneedles are used for minimally invasive medical treatments like sampling blood or subcutaneously medication delivery [56], [57]. The microneedle allows for precise localization of the medication to obtain effective absorption into the bloodstream [58]. An example of an application field for microneedles is self-monitoring of blood glucose for diabetes patients [59]. The mosquito proboscis, the mouthparts of the female mosquito that she uses to suck up blood, is reported as a source of inspiration to design advanced microneedles [60]. The mosquito's biting process and its different phases were detected electronically by Kashin and Wakely [61] and Kashin [62]. The mosquito (*Aedes albopictus* Skuse) was used as an electrical switch where electrical conduction was achieved only when there was penetration. Ramasubramanian *et al.* [63] used high-speed video imaging to study the fascicle insertion of the *Aedes aegypti* (Linnaeus) mosquito species. The female mosquito can penetrate the skin with a flexible needle that is small and flexible and can draw blood. In contrast, human-made polymeric microneedles are often not stiff enough to penetrate the skin without buckling. High-speed video images showed that the frequency of the head lateral movement decreased from 15–17 Hz in the beginning to 6 Hz towards the middle to end of the penetration. The authors presented an analytical study using a mathematical model where the fascicle is modeled as a slender column supported on an elastic foundation, subjected to non-conservative and conservative loads at the end. The authors concluded that the lateral support of the labium and the non-conservative loads help the mosquito fascicle penetrate the skin. Kong and Wu [60] studied the *A. albopictus* (Skuse) mosquito species to investigate the mechanical insertion force

of the fascicle into the skin using scanning electron microscope imaging and high-speed video imaging. Experimental results showed that in the early stages of penetration, the maxilla frequency is 10-15 Hz. This is reduced to 6-8 Hz at half depth to 3-5 Hz in the last stages of the penetration process. These maxilla frequencies comply with the results reported by Ramasubramanian *et al.* [63]. The penetration time was 10-20 s; suction time was 2-3 min; the quantity of sucked blood was about 3 mg. The measured insertion force range is 6-38  $\mu\text{N}$ , with a mean value of 16.5  $\mu\text{N}$ , three orders of magnitude smaller than the reported minimum insertion force to penetrate human skin. This is explained by the variable frequency saw-like maxillae that cut into the tissue of the skin.

Oka *et al.* [64] were first to develop a hollow microneedle inspired by the mosquito proboscis, with a jagged shape inspired by the mosquito's maxillae to ease cutting and decrease the contact area between the needle and the cutting surface. The authors manufactured a silicon microneedle with a jagged shape, reinforced with polycrystalline silicon with a length of 1 mm and an outer diameter of 85  $\mu\text{m}$ , used for trace blood tests in diabetes patients. An insertion force of 14.7 mN was found to be sufficient to penetrate the skin.

Izumi *et al.* [65] developed electrochemically etched silicon needles with cooperative motions in a parallel configuration, imitating the mosquito labium and two maxillae. Experimental results showed that cooperative motion resulted in a puncture force of 58.8 mN, compared to 205.9 mN for no motion. The authors also developed a polylactic acid (PLA) needle, considering medical applications, because PLA is biodegradable, non-brittle, and flexible, preventing possible broken pieces from remaining in the body. The PLA and silicon needles showed comparable piercing forces. Polymers like the biodegradable PLA are considered the most promising materials for microneedle fabrication due to their favorable biocompatibility [66].

Similar jagged-shaped microneedles are presented in a number of other studies, made from PLA [67], [68], silicon [69], [70], stainless steel [71], [72], and tungsten with a biocompatible coating [57]. A jagged-shaped, vibrated microneedle made from PLA is presented in [73]. Jagged-shaped vibrated microneedles that consist of multiple parallel segments are presented in a number of other studies, made from PLA [74], silicon [75], silicon with a biocompatible coating [76], photocurable epoxy resin [77], ceramic [78], and a frequency-dependent viscoelastic material [79]. Smooth-surface vibrated microneedles made from silicon [58] or titanium for a micro pumping device are presented in a number of other studies [59], [80]–[82]. Other mosquito-inspired smooth-surface microneedles are presented in a number of other studies, made from stainless steel [83], silicon [84], and shape memory polymer [85].

### B. Wasp-Inspired Steerable and Self-Propelling Needles

The ovipositor is a tube-like organ that the wasp uses for egg-laying. Sakes *et al.* [55] stated that the Hymenoptera ovipositor combines multiple buckling prevention strategies. For Hymenoptera, the ovipositor is a piercing

organ, comprising three separate pieces, capable of penetrating different materials such as wood. In order to increase the critical buckling load, the ovipositor shows amongst others the following characteristics: (1) it contains metal ions manganese and zinc that increases the bending stiffness [86], (2) the internal morphology retains the circular cross-section during penetration, this keeps the second moment of area high [55], (3) it shows barbed anchoring, which decreases the effective-length factor [60], (4) the ovipositor of Hymenoptera *Megarhyssa nortoni* (Cresson) is supported by “a longitudinal median groove flanked by series of tubercles” [87], and (5) it uses reciprocating penetration [88]. The barbed anchoring and reciprocating penetrating motion are strategies that also facilitate the penetration of the mosquito proboscis [63], [74]. Besides the buckling prevention strategies, the three longitudinal segments of the ovipositor allow for a reorientation of the ovipositor tip to a bevel-shaped tip [89], [90]. By antagonistic movements of the ovipositor segments, steering is possible. The ovipositor moves without rotation around the long center axis [91]–[93], causing less strain to the surrounding material [94]. Conventional instruments used for minimally invasive (MI) surgical interventions follow a straight-line trajectory, which reduces the planning choices of the surgeon [95]. The steering capabilities of the ovipositor of Hymenoptera make it an interesting source of inspiration for a probe capable of steering along a curved path. Neurosurgery is one field of application for steerable needles. Other fields are soft tissue applications, e.g., brachytherapy, core needle biopsy, and drug delivery. Soft tissue applications require the outer needle diameter to be below 2 mm (14 Gauge) [89], [96], [97]. Besides steerable needles, the Hymenoptera ovipositor also inspired the design of devices such as planetary and earth drills [98]. In this section, we examined some scientific publications that presented their development of steerable needles inspired by Hymenoptera.

Researchers from the Imperial College London worked on designing a steerable needle inspired by the Hymenoptera ovipositor [99]–[105]. They studied the penetration mechanism of the wasp's ovipositor, the interlocking mechanism of the ovipositor segments, and the surface texture of the ovipositor. Frasson *et al.* [95] designed a probe to access deep brain lesions through curved trajectories. The probe consists of two parallel segments with an interlocking mechanism similar to the olistheter in the Hymenoptera ovipositor, which interconnects the three segments the ovipositor consists of. The probe was produced using 3D printing and had an outer diameter of 4.4 mm. The two segments are actuated independently. When one of the segments is moved, off-axis reaction forces cause the segment to deflect in the bevel direction. The other segments follow the curved trajectory laid out by the first segment; this allows two-dimensional steering. Similar steerable probe designs consisting of two parallel needle segments were presented in a number of other studies [106]–[108]. Burrows *et al.* [109] developed a probe prototype made of a novel composite structure of alternating rigid and soft sections produced using 3D printing. The rigid sections are located at the interlocking sections and add strength to this mechanism. The soft regions are located in



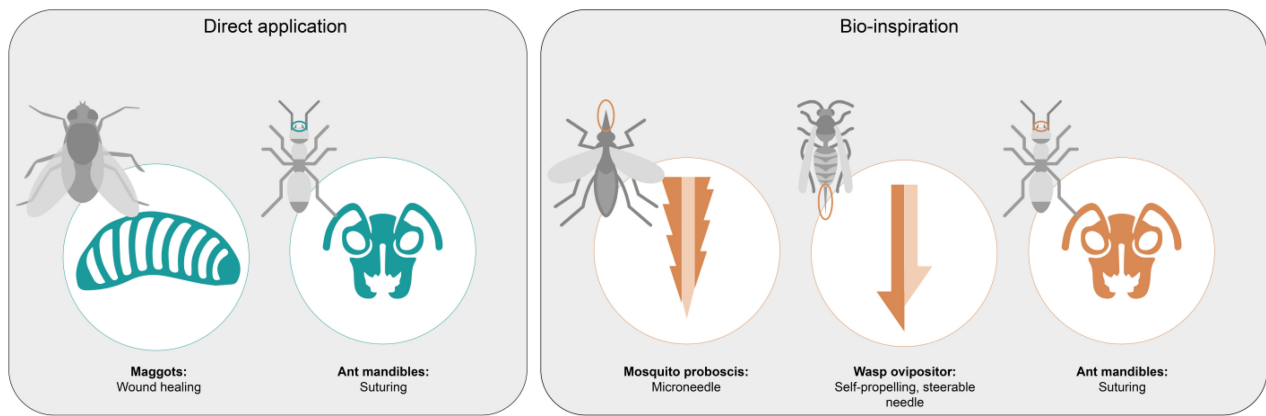


Fig. 3. Visual representation of the direct application and bio-inspiration of insects in medicine described in this review.

the regions between the rigid sections and allow the prototype to remain flexible. The needle consists of four parallel needle segments and has an outer diameter of 4 mm. Experiments in 4.5%wt gelatin showed the capability of steering in 3D with a mean positional error of 0.46 mm and an approach angle error of  $1.05^\circ$ . Similar steerable probe designs consisting of four parallel needle segments with a focus on the cross-section geometry [110], [111], on the composite body structure, on the control system for steering [94], [112], [113], on 3D steering [114], on the probe-tissue interaction [115], or on the integration with a laser Doppler system [116] have also been reported.

Sprang *et al.* [117] developed a smooth-surface four-segment prototype inspired by the Hymenoptera ovipositor. The authors stated that the ovipositor contains serrations that induce friction, which might cause unwanted tissue damage in a medical application. Therefore, they produced a multi-segment needle without gripping textures that could penetrate tissue with a low net push force. Scali *et al.* [89] decided to step away from nature with the interlocking mechanism and instead used an interlocking ring [89], [118] or a 10-mm piece of thin-walled shrinking tube [119] in combination with off-the-shelf Nitinol rods. Since no interlocking mechanism had to be produced, a tip diameter of up to 0.4 mm could be obtained. This is thinner than the other ovipositor-inspired needles in literature, where the outer diameter ranges between 4 and 12 mm [94], [95], [108], [109], [111], [113], [114]. One or two rods were actuated simultaneously, whereas the remaining stationary rods generated a friction force to compensate for the friction and cutting force of the advancing wires. Experimental evaluation of the 1.2-mm diameter prototype with an interlocking ring [89] in 4%wt gelatin showed that the needle required no external push force and allowed steering through the bevel-tip. The resultant steering curvature was  $0.000184 \text{ mm}^{-1}$ , and the deflection-to-insertion ratio was 0.0778, which is lower than previously reported steering performances [94], [108], [114], possibly due to bifurcation of the wires.

### C. Bio-Inspired Clamp

The insect family Formicidae of the order Hymenoptera is the main source of inspiration in bio-inspired clamps. Brito *et al.* [120], [121] designed a surgical clamp inspired

by the *Atta laevigata* (Smith) ant. The aim of the clamp is to make placement and removal of the clamp more efficient and less traumatic for the patient. The clamp consists of a handle structure used to open the clamp and a system in contact with the skin. The working mechanism of the surgical clamp mimics the ant mandibles: (1) an external compressive stress must be applied to the levers to open the clamp, (2) the surgical clamp can penetrate the skin due to elastic forces, (3) the clamp falls by itself after healing, which relieves the inconvenience of the clip removal process. Brito *et al.* [120] studied the mandibles of the *A. laevigata* ant to present a selection of candidate biocompatible materials for surgical clamps. The biological materials of insects' mandibles can be correlated with polymers because the mandibles are composed of waxes, polysaccharides, and proteins. Nanoindentation measurements showed that the hardness in the internal and external regions was  $0.36 \pm 0.06 \text{ GPa}$  and  $0.19 \pm 0.04 \text{ GPa}$ , respectively and the elastic modulus  $6.16 \pm 0.23 \text{ GPa}$  and  $2.74 \pm 0.44 \text{ GPa}$ , respectively. Atomic force microscopy (AFM) showed an average roughness of  $6.73 \pm 0.90 \text{ nm}$  and  $11.87 \pm 1.42 \text{ nm}$ , respectively. Zinc and manganese in the internal region justify the increase of hardness and elastic modulus in this region [86]. The authors concluded that appropriate materials for the surgical clamps include PLA, polycaprolactone, or polyglycolide acid, possibly in combination with natural polymers as chitin, collagen, or fibroin. For the handle structure, metallic biomaterials as stainless steels and titanium alloys are proposed. In another study, Brito *et al.* [121] studied the geometry and material selection of the biomimetic surgical clip using the finite element method (FEM) in ANSYS. AISI 316L and AISI 420 stainless steel were selected for the stress and strain analyses. The locations with the highest stress concentrations were determined, FEM results showed plastic deformation and possible rupture [121]. The results indicate that either the design's geometry needs optimization or the needle design should be made out of ferritic and martensitic stainless steel [121].

### V. DISCUSSION

Figure 3 shows a schematic overview of the direct application and bio-inspiration of insects used in medicine and medical engineering, respectively, described in this review.

This review also describes the direct application of maggots for wound healing. The FDA approved the medicinal maggots in 2004 and approved a preassembled version, including a netting cage in 2007 [45]. Nevertheless, treatments directly applying insects, e.g., maggot therapy, face the challenge of social approval. The poor acceptance by both patients and clinicians hinders the utilization of maggots in medicine [122]. Raising the awareness about maggot therapy and adequate psychological preparation for the patients undergoing maggot therapy might alleviate concerns about the treatment and might increase its social approval. Another method would be to extract the working principle of the maggots for medical application. During the 1930s, Livingston [123] attempted to isolate the larvae's active principle in a maggot extract. The vaccine was abandoned because of significant systemic reactions. Sherman [42] stated that someday maggot-derived products might replace live maggot larvae for wound care, e.g., antimicrobial agents capable of suppressing methicillin-resistant *Staphylococcus aureus* [124], [125]. Čerovský *et al.* [124] stated that lucifensin is assumed to be the key component that protects maggots in the infectious environment of the wound during MDT and is believed to be effective against pathogenic elements in the wound such as methicillin-resistant *Staphylococcus aureus*. The authors reported the presence of lucifensin in the gut, salivary glands, fat body, and hemolymph of the *L. sericata* larvae. Lucifensin was also detected in the washes of maggot larvae removed from the wound of a diabetic patient. Mass spectrometry and reversed-phase high-performance liquid chromatography showed the anti-*Micrococcus luteus* (Schroeter) activity of lucifensin. Andersen *et al.* [126] tried to identify the antibacterial mechanism in maggots and to purify the components that could be used in medicine. The authors stated they did not find an obvious protein with antimicrobial activity, so the authors could not pinpoint lucifensin as the key component that is effective against pathogenic elements. This is in contrast with the results of Čerovský *et al.* [124]. Lucifensin showed to be active against Gram-positive bacteria, whereas no antimicrobial activity towards Gram-negative bacteria was shown. The authors stated that in future studies, the expression of lucifensin at different maggot instars should be validated in order to evaluate whether lucifensin is responsible for the antimicrobial activity in maggot excretions and secretions. In 1995, Morgan [30] already stated an opportunity to develop genetically engineered flies bred in a sterile environment. Linger *et al.* [127] presented a novel concept that combines MDT benefits with genetic engineering to promote wound healing. The authors investigated human platelet-derived growth factor-BB as a potential treatment for non-healing wounds, as it is known to promote wound healing [128]. The authors produced genetically modified *L. sericata* larvae that secrete platelet-derived growth factor-BB at a detectable level in maggot ASE. The effector genes could be other growth factors such as lucifensin. This would indicate that ASE would be active against Gram-positive bacteria [124].

Mosquitoes and wasps are both capable of penetrating solid substrates. In the scientific literature, two common biological

features were described that facilitate the needle insertion, the serrated needle segments and the reciprocating motion of the needle segments [129]. Besides the mosquito proboscis and the wasp ovipositor, other parts of insects used as inspiration for the design of medical needles are the mouthpart of the tsetse fly [130], [131], the mouthpart of Cicadellidae [132], the spine of the caterpillar [133], and stings of honey bees and paperwasps [134]. For future research, it is interesting to look into the mechanical differences of the medical needles inspired by these different insects. Besides the insects and insect-inspired products described in this review, other insects and insect-derived products are used in folk healing that might have potential in conventional medicine. Cherniack [16] wrote a review of the use of insects and insect-derived products in folk healing. Besides maggot therapy, the author included honey treatment, the use of royal jelly, and the application of bee and ant venom and blister beetle-derived cantharidin. Honey is used to heal wounds or treat burns [135]. Royal jelly is used to treat postmenopausal symptoms [136]. Bee and ant venom is used to treat swollen joints in patients with rheumatoid arthritis [137]. Blister beetle-derived cantharidin is used to treat warts and molluscum [138]. The *Decticus verrucivorus* (Linnaeus), also called the wart-biter, has gotten its name from the old Swedish practice of allowing the insect to bite warts from the skin [139]. In Chinese medicine, chemotherapy is combined with insect therapy for non-surgical liver tumor treatment [140]. Sodium cantharidinate and vitamin B6 injections and cantharides capsules are common insect Chinese medicine [140] that contain extractions from the blister beetle (*Mylabris variabilis* Pallas) [141]. The egg cases of the praying mantis (*Mantis* Linnaeus) are used in traditional Chinese medicine to treat incontinence and frequent urination [142]. Furthermore, Chinese medicine also utilizes products derived from other animal species and herbs besides insects [140]. Another insect-inspired product described in the scientific literature is the physical-based bactericidal surface [143]. Ishwarya *et al.* [144] described the use of the seed extract of *Pedaliium murex* Linn (family: Pedaliaceae) to produce silver nanoparticles. The study showed that the produced silver nanoparticles combined with Gram-positive and Gram-negative bacteria result in a larger quantity of dead bacteria than the control sample. Furthermore, experiments on mosquito larvae showed the larvicidal potential of the silver nanoparticles. Jaggessar *et al.* [143] reviewed both the natural and bio-inspired antibacterial surfaces. Nanotextured surfaces were produced inspired by the dragonfly wings, cicada wings, and butterfly wings. The surfaces produced showed varying bactericidal efficiencies [143]. Hence, additional research should investigate how to engineer a surface pattern that incorporates the best features of nano-surfaces found on insect wings. These surfaces are a helpful addition for medical implants with an associated risk of bacterial infection.

For insect-inspired medical instruments, one of the main challenges is the complexity of the insect structures found in nature. Structures such as the interlocking mechanism found in the wasp ovipositor are not possible to produce using the current microfabrication techniques [55]. This and other challenges, such as the production of biocompatible materials, might explain why many insect mechanisms are not yet



applied in the development of innovative biomedical instruments. With this review, we would like to propose a new field of research called “entomomedical engineering.” We define entomomedical engineering as the branch of engineering that uses insects either directly or as a source of inspiration to design and develop innovative medical strategies. The diversity of insects provides a great source of inspiration for us to evolve innovative medical treatments and instruments.

## VI. CONCLUSION

This work provides an overview of the state of the art in the scientific literature describing the direct application or the bio-inspiration of insects used in medical engineering. The goal was to analyze the different insects used directly or as inspiration source. This research field is what we call entomomedical engineering, i.e., an innovative branch of engineering that uses insects either directly or as a source of inspiration for innovative medical technologies. We discussed a total of 93 relevant scientific articles found in the Scopus database published over the last 20 years (2000-2020). Medical treatments directly applying insects face the challenge of social approval by both patients and clinicians. Raising awareness about insect application in medicine or extracting the insect’s working principle might increase its social approval. One of the main challenges for insect-inspired medical instruments is the complexity of the insect structures found in nature. Increasing the development of microfabrication techniques might increase the clinical application of insect-inspired medical instruments. Novel medical instruments and treatments can be developed by exploring the field of entomomedical engineering.

## REFERENCES

- [1] O. Dangles and J. Casas, “Ecosystem services provided by insects for achieving sustainable development goals,” *Ecosyst. Serv.*, vol. 35, pp. 109–115, Feb. 2019.
- [2] M. Chagnon, D. Kreutzweiser, E. A. Mitchell, C. A. Morrissey, D. A. Noome, and J. P. Van der Sluijs, “Risks of large-scale use of systemic insecticides to ecosystem functioning and services,” *Environ. Sci. Pollut. Res.*, vol. 22, no. 1, pp. 119–134, 2015.
- [3] B. C. Lister and A. Garcia, “Climate-driven declines in arthropod abundance restructure a rainforest food web,” *Proc. Nat. Acad. Sci.*, vol. 115, no. 44, pp. E10397–E10406, 2018.
- [4] L. H. Yang and C. Gratton, “Insects as drivers of ecosystem processes,” *Current Opinion Insect Sci.*, vol. 2, pp. 26–32, Aug. 2014.
- [5] C. Kremen and R. Chaplin-Kramer, “Insects as providers of ecosystem services: Crop pollination and pest control,” in *Proc. Insect Conserv. Biol. Proc. Roy. Entomol. Soc. 23rd Symp.*, 2007, pp. 349–382.
- [6] E. Öckinger and H. G. Smith, “Semi-natural grasslands as population sources for pollinating insects in agricultural landscapes,” *J. Appl. Ecol.*, vol. 44, no. 1, pp. 50–59, 2007.
- [7] P. Jouquet, J. Dauber, J. Lagerlöf, P. Lavelle, and M. Lepage, “Soil invertebrates as ecosystem engineers: Intended and accidental effects on soil and feedback loops,” *Appl. Soil Ecol.*, vol. 32, no. 2, pp. 153–164, 2006.
- [8] T. G. Benton, D. M. Bryant, L. Cole, and H. Q. Crick, “Linking agricultural practice to insect and bird populations: A historical study over three decades,” *J. Appl. Ecol.*, vol. 39, no. 4, pp. 673–687, 2002.
- [9] S. N. Gorb, M. Sinha, A. Peressadko, K. A. Daltorio, and R. D. Quinn, “Insects did it first: A micropatterned adhesive tape for robotic applications,” *Bioinspiration Biomimetics*, vol. 2, no. 4, p. S117–S125, 2007.
- [10] D. Floreano, J.-C. Zufferey, M. V. Srinivasan, and C. Ellington, *Flying Insects and Robots*. Berlin, Germany: Springer-Verlag, 2009.
- [11] D. Grimaldi and M. Engel, *Evolution of the Insects*, 15 ed. Cambridge, U.K.: Cambridge Univ. Press, 2005.
- [12] E. M. Costa-Neto, “Entomotherapy, or the medicinal use of insects,” *J. Ethnobiol.*, vol. 25, no. 1, pp. 93–114, 2005.
- [13] N. Rastogi, “Provisioning services from ants: Food and pharmaceuticals,” *Asian Myrmecol.*, vol. 4, no. 1, pp. 103–120, 2011.
- [14] L. J. Rather, M. F. Ansari, and Q. Li, “Recent advances in the insect natural product chemistry: Structural diversity and their applications,” in *Natural Materials and Products from Insects: Chemistry and Applications*. Cham, Switzerland: Springer, 2020, pp. 67–94.
- [15] R. W. Pemberton, “Insects and other arthropods used as drugs in Korean traditional medicine,” *J. Ethnopharmacol.*, vol. 65, no. 3, pp. 207–216, 1999.
- [16] E. P. Cherniack, “Bugs as drugs, part 1: Insects: The ‘new’ alternative medicine for the 21st century?” *Altern. Med. Rev.*, vol. 15, no. 2, pp. 124–135, 2010.
- [17] G. Vogel, *Where Have All the Insects Gone?* Washington, DC, USA: Amer. Assoc. Adv. Sci., 2017.
- [18] C. A. Hallmann *et al.*, “More than 75 percent decline over 27 years in total flying insect biomass in protected areas,” *PLoS ONE*, vol. 12, no. 10, 2017, Art. no. e0185809.
- [19] R. van Klink, D. E. Bowler, K. B. Gongalsky, A. B. Swengel, A. Gentile, and J. M. Chase, “Meta-analysis reveals declines in terrestrial but increases in freshwater insect abundances,” *Science*, vol. 368, no. 6489, pp. 417–420, 2020.
- [20] G. A. Montgomery *et al.*, “Is the insect apocalypse upon us? How to find out,” *Biol. Conserv.*, vol. 241, Jan. 2020, Art. no. 108327.
- [21] M. L. Forister, E. M. Pelton, and S. H. Black, “Declines in insect abundance and diversity: We know enough to act now,” *Conserv. Sci. Pract.*, vol. 1, no. 8, p. e80, 2019.
- [22] F. Sánchez-Bayo and K. A. G. Wyckhuys, “Worldwide decline of the entomofauna: A review of its drivers,” *Biol. Conserv.*, vol. 232, pp. 8–27, Apr. 2019.
- [23] R. F. Pereira and P. J. Bartolo, “Traditional therapies for skin wound healing,” *Adv. Wound Care*, vol. 5, no. 5, pp. 208–229, 2016.
- [24] U. Wollina, K. Karte, C. Herold, and A. Looks, “Biosurgery in wound healing—the renaissance of maggot therapy,” *J. Eur. Acad. Dermatol. Venereol.*, vol. 14, no. 4, pp. 285–289, 2000.
- [25] R. A. Sherman, K. Y. Mumcuoglu, M. Grassberger, and T. I. Tantawi, “Maggot therapy,” in *Biotherapy-History, Principles and Practice*, Dordrecht, The Netherlands: Springer, 2013, pp. 5–29.
- [26] R. A. Sherman, “Maggot versus conservative debridement therapy for the treatment of pressure ulcers,” *Wound Repair Regen.*, vol. 10, no. 4, pp. 208–214, 2002.
- [27] J. Wayman, V. Nirojogi, A. Walker, A. Sowinski, and M. A. Walker, “The cost effectiveness of larval therapy in venous ulcers,” *J. Tissue Viability*, vol. 10, no. 3, pp. 91–94, 2000.
- [28] C. Wilarusmee *et al.*, “Maggot therapy for chronic ulcer: A retrospective cohort and a meta-analysis,” *Asian J. Surg.*, vol. 37, no. 3, pp. 138–147, 2014.
- [29] F. Zumpt, *Myiasis in Man and Animals in the Old World: A Textbook for Physicians, Veterinarians and Zoologists*. London, U.K.: Butterworths, 1965.
- [30] D. Morgan, “Myiasis: The rise and fall of maggot therapy,” *J. Tissue Viability*, vol. 5, no. 2, pp. 43–51, 1995.
- [31] S. Thomas, M. Jones, S. Shutler, and S. Jones, “Using larvae in modern wound management,” *J. Wound Care*, vol. 5, no. 2, pp. 60–69, 1996.
- [32] R. A. Sherman and E. A. Pechter, “Maggot therapy: A review of the therapeutic applications of fly larvae in human medicine, especially for treating osteomyelitis,” *Med. Vet. Entomol.*, vol. 2, no. 3, pp. 225–230, 1988.
- [33] L. M. Vistnes, R. Lee, and G. A. Ksander, “Proteolytic activity of blowfly larvae secretions in experimental burns,” *Surgery*, vol. 90, no. 5, pp. 835–841, 1981.
- [34] S. E. Ziffren, H. E. Heist, S. C. May, and N. A. Womack, “The secretion of collagenase by maggots and its implication,” *Ann. Surg.*, vol. 138, no. 6, p. 932, 1953.
- [35] H. I. Goldstein, “Maggots in the treatment of wound and bone infections,” *J. Bone Joint Surg.*, vol. 13, no. 3, pp. 476–478, 1931.
- [36] W. S. Baer, “The treatment of chronic osteomyelitis with the maggot (larva of the blow fly),” *J. Bone Joint Surg.*, vol. 13, no. 3, pp. 438–475, 1931.
- [37] E. A. Pechter and R. A. Sherman, “Maggot therapy: The surgical metamorphosis,” *Plast. Reconstr. Surg.*, vol. 72, no. 4, pp. 567–570, 1983.
- [38] R. A. Sherman, M. Hall, and S. Thomas, “Medicinal maggots: An ancient remedy for some contemporary afflictions,” *Ann. Rev. Entomol.*, vol. 45, no. 1, pp. 55–81, 2000.

- [39] M. Wainwright, "Maggot therapy—A backwater in the fight against bacterial infection," *Pharm. Hist.*, vol. 30, no. 1, pp. 19–26, 1988.
- [40] Y. Nigam, A. Bexfield, S. Thomas, and N. A. Ratcliffe, "Maggot therapy: The science and implication for CAM part I—History and bacterial resistance," *Evidence Based Complement. Altern. Med.*, vol. 3, no. 2, pp. 223–227, 2006.
- [41] Y. Nigam, A. Bexfield, S. Thomas, and N. A. Ratcliffe, "Maggot therapy: The science and implication for CAM part II—Maggots combat infection," *Evidence Based Complement. Altern. Med.*, vol. 3, no. 3, pp. 303–308, 2006.
- [42] R. A. Sherman, "Maggot therapy takes us back to the future of wound care: New and improved maggot therapy for the 21st century," *J. Diabetes Sci. Technol.*, vol. 3, no. 2, pp. 336–344, 2009.
- [43] R. A. Sherman, "Mechanisms of maggot-induced wound healing: What do we know, and where do we go from here?" *Evidence Based Complement. Altern. Med.*, vol. 2014, Mar. 2014, Art. no. 592419.
- [44] N. Andersson, C. P. Da Sousa, and S. Paredes, "Social cost of land mines in four countries: Afghanistan, Bosnia, Cambodia, and Mozambique," *BMJ*, vol. 311, no. 7007, pp. 718–721, 1995.
- [45] FDA. 510(k) Premarket Notification, Medical Maggots, K033391. Accessed: Dec. 13, 2019. [Online]. Available: <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm?ID=13466>
- [46] EWMA. *How Do Maggots Operate*. Accessed: Dec. 13, 2019. [Online]. Available: <https://ewma.org/nl/ewma-conference-dutch/2017-nl/scientific/workshops/how-do-maggots-operate/>
- [47] O. von Beckerath, S. Kanya, G. Gäbel, K. Kröger, and B. Juntermanns, "Use of maggot debridement therapy in hospitalised patients in Germany," *Int. Wound J.*, vol. 17, no. 1, pp. 10–15, 2020.
- [48] D. Mackenzie, "The history of sutures," *Med. Hist.*, vol. 17, no. 2, pp. 158–168, 1973.
- [49] G. J. Lockhart, "Ants and other great medicines," unpublished.
- [50] F. S. Haddad, "Suturing methods and materials with special emphasis on the jaws of giant ants: An old-new surgical instrument," *Lebanese Med. J.*, vol. 103, no. 359, pp. 1–4, 2010.
- [51] A. Scierski, "From ant to stapler-100 years of mechanical suturing in surgery," *Wideochirurgia i Inne Techniki Malo Inwazyjne*, vol. 5, no. 2, p. 76, 2010.
- [52] J. Schiappa and R. V. Hee, "From ants to staples: History and ideas concerning suturing techniques," *Acta Chirurgica Belgica*, vol. 112, no. 5, pp. 395–402, 2012.
- [53] C. Iavazzo, M. Papakirtsis, M. Karamanou, F. Ntziora, and G. Androutsos, "Ant mandibles as staples in the era of the greek patriot ioannis makriyannis (1797–1864)," *Acta Medico Historica Adriatica*, vol. 11, no. 2, pp. 359–364, 2013.
- [54] P. Kripouri and D. Filippou, "Sutures in antiquity," *J. Surg.*, vol. 10, pp. 2575–9760, Feb. 2018.
- [55] A. Sakes, D. Dodou, and P. Breedveld, "Buckling prevention strategies in nature as inspiration for improving percutaneous instruments: A review," *Bioinspiration Biomimetics*, vol. 11, no. 2, 2016, Art. no. 021001.
- [56] G. E. Gattiker, K. V. Kaler, and M. P. Mintchev, "Electronic Mosquito: Designing a semi-invasive Microsystem for blood sampling, analysis and drug delivery applications," *Microsyst. Technol.*, vol. 12, no. 1–2, pp. 44–51, 2005.
- [57] T. Tanaka, T. Takahashi, M. Suzuki, and S. Aoyagi, "Development of minimally invasive microneedle made of tungsten—Sharpening through electrochemical etching and hole processing for drawing up liquid using excimer laser —," *J. Robot. Mechatron.*, vol. 25, no. 4, pp. 755–761, 2013.
- [58] M. Yang and J. D. Zahn, "Microneedle insertion force reduction using vibratory actuation," *Biomed. Microdevices*, vol. 6, no. 3, pp. 177–182, 2004.
- [59] K. Tsuchiya, N. Nakanishi, Y. Uetsuji, and E. Nakamachi, "Development of blood extraction system for health monitoring system," *Biomed. Microdevices*, vol. 7, no. 4, pp. 347–353, 2005.
- [60] X. Q. Kong and C. W. Wu, "Mosquito proboscis: An elegant biomicro-electromechanical system," *Phys. Rev. E, Stat. Phys. Plasmas Fluids Relat. Interdiscip. Top.*, vol. 82, no. 1, 2010, Art. no. 011910.
- [61] P. Kashin and H. G. Wakeley, "An insect 'bitometer,'" *Nature*, vol. 208, no. 5009, pp. 462–464, 1965.
- [62] P. Kashin, "Electronic recording of the mosquito bite," *J. Insect Physiol.*, vol. 12, no. 3, pp. 281–286, 1966.
- [63] M. K. Ramasubramanian, O. M. Barham, and V. Swaminathan, "Mechanics of a mosquito bite with applications to microneedle design," *Bioinspiration Biomimetics*, vol. 3, no. 4, 2008, Art. no. 046001.
- [64] K. Oka, S. Aoyagi, Y. Arai, Y. Isono, G. Hashiguchi, and H. Fujita, "Fabrication of a micro needle for a trace blood test," *Sens. Actuators A, Phys.*, vol. 97–98, pp. 478–485, Apr. 2002.
- [65] H. Izumi, M. Suzuki, S. Aoyagi, and T. Kanzaki, "Realistic imitation of mosquito's proboscis: Electrochemically etched sharp and jagged needles and their cooperative inserting motion," *Sens. Actuators A, Phys.*, vol. 165, no. 1, pp. 115–123, 2011.
- [66] G. Ma and C. Wu, "Microneedle, bio-microneedle and bio-inspired microneedle: A review," *J. Control. Release*, vol. 251, pp. 11–23, Apr. 2017.
- [67] S. Aoyagi, H. Izumi, T. Aoki, and M. Fukuda, "Development of a micro lancet needle made of biodegradable polymer for low-invasive medical treatment," in *Proc. Transducers*, vol. 2, 2005, pp. 1195–1198.
- [68] S. Aoyagi, H. Izumi, Y. Isono, K. Makihira, and M. Fukuda, "Biodegradable polymer needle having a trench for collecting blood by capillary force," in *Proc. 19th IEEE Int. Conf. Micro Electro Mech. Syst.*, 2006, pp. 450–453.
- [69] H. Izumi and S. Aoyagi, "Novel fabrication method for long silicon microneedles with three-dimensional sharp tips and complicated shank shapes by isotropic dry etching," *IEEEJ Trans. Elect. Electron. Eng.*, vol. 2, no. 3, pp. 328–334, 2007.
- [70] H. Izumi, T. Okamoto, M. Suzuki, and S. Aoyagi, "Development of a silicon microneedle with three-dimensional sharp tip by electrochemical etching," *IEEEJ Trans. Sens. Micromach.*, vol. 129, no. 11, pp. 373–379, 2009.
- [71] Y. Hara, M. Yamada, C. Tatsukawa, T. Takahashi, M. Suzuki, and S. Aoyagi, "Fabrication of stainless steel microneedle with laser-cut sharp tip and its penetration and blood sampling performance," *Int. J. Autom. Technol.*, vol. 10, no. 6, pp. 950–957, 2016.
- [72] Y. Hara, M. Yamada, C. Tatsukawa, T. Takahashi, M. Suzuki, and S. Aoyagi, "Laser fabrication of jagged-shaped stainless steel microneedle imitating mosquito's maxilla," *Int. J. Autom. Technol.*, vol. 10, no. 6, pp. 958–964, 2016.
- [73] S. Aoyagi, H. Izumi, and M. Fukuda, "Biodegradable polymer needle with various tip angles and effect of vibration and surface tension on easy insertion," in *Proc. IEEE 20th Int. Conf. Micro Electro Mech. Syst. (MEMS)*, 2007, pp. 397–400.
- [74] S. Aoyagi, H. Izumi, and M. Fukuda, "Biodegradable polymer needle with various tip angles and consideration on insertion mechanism of mosquito's proboscis," *Sens. Actuators A, Phys.*, vol. 143, no. 1, pp. 20–28, 2008.
- [75] S. Aoyagi *et al.*, "Equivalent negative stiffness mechanism using three bundled needles inspired by mosquito for achieving easy insertion," in *Proc. IEEE/RSJ Int. Conf. Intell. Robots Syst.*, 2012, pp. 2295–2300.
- [76] H. Izumi *et al.*, "Combined harpoonlike jagged microneedles imitating mosquito's proboscis and its insertion experiment with vibration," *IEEEJ Trans. Elect. Electron. Eng.*, vol. 3, no. 4, pp. 425–431, 2008.
- [77] M. Suzuki, T. Sawa, Y. Terada, T. Takahashi, and S. Aoyagi, "Fabrication of microneedles precisely imitating mosquito's proboscis by nanoscale tree dimensional laser lithography and its characterization," in *Proc. Transducers 18th Int. Conf. Solid-State Sens. Actuat. Microsyst. (TRANSDUCERS)*, Anchorage, AK, USA, 2015, pp. 121–124.
- [78] S. D. Gittard, R. J. Narayan, A. Ovsianikov, and B. N. Chichkov, "Rapid prototyping of biomimetic structures: Fabrication of mosquito-like microneedles by two-photon polymerization," in *MRS Online Proceedings Library Archive*, vol. 1239. Cambridge, U.K.: Cambridge Univ. Press, 2009.
- [79] D. Gurera, B. Bhushan, and N. Kumar, "Lessons from mosquitoes' painless piercing," *J. Mech. Behav. Biomed. Mater.*, vol. 84, pp. 178–187, Aug. 2018.
- [80] K. Tsuchiya, A. Morishima, A. Takamata, Y. Uetsuji, and E. Nakamachi, "Development of valve-less tube-type micropump with PZT actuator," in *Proc. Biomed. Appl. Micro Nanoeng. IV Complex Syst.*, vol. 7270, 2008, Art. no. 72700E.
- [81] K. Tsuchiya, S. Jinnin, H. Yamamoto, Y. Uetsuji, and E. Nakamachi, "Design and development of a biocompatible painless microneedle by the ion sputtering deposition method," *Precis. Eng.*, vol. 34, no. 3, pp. 461–466, 2010.
- [82] K. Tsuchiya, "The painless injection tube: From bio-mimetic technology to medical engineering," in *Thought-Evoking Approaches in Engineering Problems*. Cham, Switzerland: Springer Int., 2014, pp. 71–94.
- [83] A. Oki *et al.*, "Healthcare chip for checking health condition from analysis of trace blood collected by painless needle," *Jpn. J. Appl. Phys.*, vol. 42, no. 6R, p. 3722, 2003.

- [84] R. Sharaf, P. Aggarwal, K. V. I. S. Kaler, and W. Badawy, "On the design of an electronic mosquito: design and analysis of the micro-needle," in *Proc. Int. Conf. MEMS NANO Smart Syst.*, Banff, AB, Canada, 2003, pp. 32–35.
- [85] A. J. Shoffstall *et al.*, "A mosquito inspired strategy to implant microprobes into the brain," *Sci. Rep.*, vol. 8, no. 1, pp. 1–10, 2018.
- [86] D. L. Quicke, P. Wyeth, J. D. Fawke, H. H. Basibuyuk, and J. F. V. Vincent, "Manganese and zinc in the ovipositors and mandibles of hymenopterous insects," *Zool. J. Linnean Soc.*, vol. 124, no. 4, pp. 387–396, 1998.
- [87] L. Vilhelmsen, N. Isidoro, R. Romani, H. H. Basibuyuk, and D. L. J. Quicke, "Host location and oviposition in a basal group of parasitic wasps: The subgenual organ, ovipositor apparatus and associated structures in the Orussidae (Hymenoptera, Insecta)," *Zoomorphology*, vol. 121, no. 2, pp. 63–84, 2001.
- [88] H. R. Hermann, "Sting autotomy, a defensive mechanism in certain social Hymenoptera," *Insectes Sociaux*, vol. 18, no. 2, pp. 111–120, 1971.
- [89] M. Scali, T. Pusch, P. Breedveld, and D. Dodou, "Ovipositor-inspired steerable needle: Design and preliminary experimental evaluation," *Bioinspiration Biomimetics*, vol. 13, no. 1, 2017, Art. no. 016006.
- [90] D. L. J. Quicke and M. G. Fitton, "Ovipositor steering mechanisms in parasitic wasps of the families Gasteruptionidae and Aulacidae (Hymenoptera)," *Proc. Roy. Soc. London B, Biol. Sci.*, vol. 261, no. 1360, pp. 99–103, 1995.
- [91] D. S. Minhas, J. A. Engh, M. M. Fenske, and C. N. Riviere, "Modeling of needle steering via duty-cycled spinning," in *Proc. 29th Annu. Int. Conf. IEEE Eng. Med. Biol. Soc.*, Lyon, France, 2007, pp. 2756–2759.
- [92] D. Minhas, J. A. Engh, and C. N. Riviere, "Testing of neurosurgical needle steering via duty-cycled spinning in brain tissue in vitro," in *Proc. Annu. Int. Conf. IEEE Eng. Med. Biol. Soc.*, Minneapolis, MN, USA, 2009, pp. 258–261.
- [93] N. A. Wood, K. Shahrou, M. C. Ost, and C. N. Riviere, "Needle steering system using duty-cycled rotation for percutaneous kidney access," in *Proc. Annu. Int. Conf. IEEE Eng. Med. Biol.*, 2010, pp. 5432–5435.
- [94] S. Y. Ko, L. Frasson, and F. R. Y. Baena, "Closed-loop planar motion control of a steerable probe with a 'programmable bevel' inspired by nature," *IEEE Trans. Robot.*, vol. 27, no. 5, pp. 970–983, Oct. 2011.
- [95] L. Frasson, S. Ko, A. Turner, T. Parittotokkaporn, J. F. Vincent, and F. R. Y. Baena, "STING: A soft-tissue intervention and neurosurgical guide to access deep brain lesions through curved trajectories," *Proc. Inst. Mech. Eng. H, J. Eng. Med.*, vol. 224, no. 6, pp. 775–788, 2010.
- [96] G. Gherardi, *Fine-Needle Biopsy of Superficial and Deep Masses: Interventional Approach and Interpretation Methodology by Pattern Recognition*. Milan, Italy: Springer-Verlag, 2010.
- [97] T. Podder *et al.*, "Vivo motion and force measurement of surgical needle intervention during prostate brachytherapy," *Med. Phys.*, vol. 33, no. 8, pp. 2915–2922, 2006.
- [98] Y. Gao, A. Ellery, M. Jaddou, and J. Vincent, "Deployable wood wasp drill for planetary subsurface sampling," in *Proc. IEEE Aerosp. Conf.*, Big Sky, MT, USA, 2006, p. 8.
- [99] L. Frasson *et al.*, "Biologically inspired microtexturing: Investigation into the surface topography of next-generation neurosurgical probes," in *Proc. 30th Annu. Int. Conf. IEEE Eng. Med. Biol. Soc.*, 2008, pp. 5611–5614.
- [100] A. Schneider, L. Frasson, T. Parittotokkaporn, F. R. Y. Baena, B. Davies, and S. Huq, "Microfabrication of components for a novel biomimetic neurological endoscope," in *Proc. 4th Int. Conf. Multi-Mater. Micro Manuf.*, 2008, pp. 9–11.
- [101] L. Frasson, T. Parittotokkaporn, B. L. Davies, and F. R. Y. Baena, "Early developments of a novel smart actuator inspired by nature," *Int. J. Intell. Syst. Technol. Appl.*, vol. 8, nos. 1–4, pp. 409–422, 2010.
- [102] L. Frasson, S. Reina, B. Davies, and F. R. Y. Baena, "Design optimisation of a biologically inspired multi-part probe for soft tissue surgery," in *Proc. World Congr. Med. Phys. Biomed. Eng.*, Munich, Germany, Sep. 2009, pp. 307–310.
- [103] T. Parittotokkaporn *et al.*, "Soft tissue traversal with zero net force: Feasibility study of a biologically inspired design based on reciprocal motion," in *Proc. IEEE Int. Conf. Robot. Biomimetics*, Bangkok, Thailand, 2009, pp. 80–85.
- [104] A. Schneider, L. Frasson, T. Parittotokkaporn, F. M. R. Y. Baena, B. L. Davies, and S. E. Huq, "Biomimetic microtexturing for neurosurgical probe surfaces to influence tribological characteristics during tissue penetration," *Microelectron. Eng.*, vol. 86, nos. 4–6, pp. 1515–1517, 2009.
- [105] T. Parittotokkaporn, L. Frasson, A. Schneider, B. L. Davies, P. Degenair, and F. R. Y. Baena, "Insertion experiments of a biologically inspired microtextured and multi-part probe based on reciprocal motion," in *Proc. Annu. Int. Conf. IEEE Eng. Med. Biol.*, Buenos Aires, Argentina, 2010, pp. 3190–3193.
- [106] S. Y. Ko, B. L. Davies, and F. R. Y. Baena, "Two-dimensional needle steering with a 'programmable bevel' inspired by nature: Modeling preliminaries," in *Proc. IEEE/RSJ Int. Conf. Intell. Robots Syst.*, Taipei, Taiwan, 2010, pp. 2319–2324.
- [107] L. Frasson, F. Ferroni, S. Y. Ko, G. Dogangil, and F. R. Y. Baena, "Experimental evaluation of a novel steerable probe with a programmable bevel tip inspired by nature," *J. Robot. Surg.*, vol. 6, no. 3, pp. 189–197, 2012.
- [108] S. Y. Ko and F. R. Y. Baena, "Toward a miniaturized needle steering system with path planning for obstacle avoidance," *IEEE Trans. Biomed. Eng.*, vol. 60, no. 4, pp. 910–917, Apr. 2013.
- [109] C. Burrows, F. Liu, A. Leibinger, R. Secoli, and F. R. Y. Baena, "Multi-target planar needle steering with a bio-inspired needle design," in *Advances in Italian Mechanism Science*. Cham, Switzerland: Springer Int., 2017, pp. 51–60.
- [110] L. Frasson, J. Neubert, S. Reina, M. Oldfield, B. L. Davies, and F. R. Y. Baena, "Development and validation of a numerical model for cross-section optimization of a multi-part probe for soft tissue intervention," in *Proc. Annu. Int. Conf. IEEE Eng. Med. Biol.*, 2010, pp. 3202–3205.
- [111] A. Leibinger, M. Oldfield, and F. R. Y. Baena, "Multi-objective design optimization for a steerable needle for soft tissue surgery," in *Proc. 15th Int. Conf. Biomed. Eng.*, 2014, pp. 420–423.
- [112] R. Secoli and F. R. Y. Baena, "Adaptive path-following control for bio-inspired steerable needles," in *Proc. 6th IEEE Int. Conf. Biomed. Robot. Biomechanics (BioRob)*, Singapore, 2016, pp. 87–93.
- [113] R. Secoli and F. Rodriguez, "Experimental validation of curvature tracking with a programmable bevel-tip steerable needle," in *Proc. Int. Symp. Med. Robot. (ISMR)*, 2018, pp. 1–6.
- [114] C. Burrows, R. Secoli, and F. R. Y. Baena, "Experimental characterisation of a biologically inspired 3D steering needle," in *Proc. 13th Int. Conf. Control Autom. Syst. (ICCAS)*, 2013, pp. 1252–1257.
- [115] A. Leibinger, M. J. Oldfield, and F. R. Y. Baena, "Minimally disruptive needle insertion: A biologically inspired solution," *Interface Focus*, vol. 6, no. 3, 2016, Art. no. 20150107.
- [116] V. Virdyawan, M. Oldfield, and F. R. Y. Baena, "Laser Doppler sensing for blood vessel detection with a biologically inspired steerable needle," *Bioinspir. Biomimetics*, vol. 13, no. 2, 2018, Art. no. 026009.
- [117] T. Sprang, P. Breedveld, and D. Dodou, "Wasp-inspired needle insertion with low net push force," in *Proc. Conf. Biomimetic Biohybrid Syst.*, 2016, pp. 307–318.
- [118] M. Scali, D. Kreeft, P. Breedveld, and D. Dodou, "Design and evaluation of a wasp-inspired steerable needle," in *Proc. SPIE Bioinspiration Biomimetics Bioreplication*, vol. 10162. Portland, OR, USA, Apr. 2017, Art. no. 1016207.
- [119] M. Scali, P. Breedveld, and D. Dodou, "Experimental evaluation of a self-propelling bio-inspired needle in single- and multi-layered phantoms," *Sci. Rep.*, vol. 9, no. 1, pp. 1–13, 2019.
- [120] T. O. Brito, B. B. D. Santos, L. S. Araújo, L. H. De Almeida, and M. F. Da Costa, "Analysis of biomimetic surgical clip using finite element modeling for geometry improvement and biomaterials selection," in *Proc. 3rd Pan Amer. Mater. Congr.*, 2017, pp. 3–9.
- [121] T. O. Brito, A. Elzubair, L. S. Araújo, S. A. D. S. Camargo, J. L. P. Souza, and L. H. Almeida, "Characterization of the mandible *Atta laevigata* and the bioinspiration for the development of a biomimetic surgical clamp," *Mater. Res.*, vol. 20, no. 6, pp. 1525–1533, 2017.
- [122] D. C. Chan, D. H. Fong, J. Y. Leung, N. G. Patil, and G. K. K. Leung, "Maggot debridement therapy in chronic wound care," *Hong Kong Med. J.*, vol. 13, no. 5, pp. 382–386, 2007.
- [123] S. Livingston, "The therapeutic active principle of maggots: With a description of its clinical application in 567 cases," *J. Bone Joint Surg.*, vol. 18, no. 3, pp. 751–756, 1936.
- [124] V. Čefovský, J. Žďárek, V. Fučík, L. Monincová, Z. Voburka, and R. Bém, "Lucifensin, the long-sought antimicrobial factor of medicinal maggots of the blowfly *Lucilia sericata*," *Cell. Mol. Life Sci.*, vol. 67, no. 3, pp. 455–466, 2010.
- [125] M. J. Van der Plas *et al.*, "Maggot excretions/secretions are differentially effective against biofilms of *Staphylococcus aureus* and *Pseudomonas aeruginosa*," *J. Antimicrobial Chemother.*, vol. 61, no. 1, pp. 117–122, 2008.

- [126] A. S. Andersen *et al.*, "A novel approach to the antimicrobial activity of maggot debridement therapy," *J. Antimicrobial Chemother.*, vol. 65, no. 8, pp. 1646–1654, 2010.
- [127] R. J. Linger *et al.*, "Towards next generation maggot debridement therapy: Transgenic *Lucilia sericata* larvae that produce and secrete a human growth factor," *BMC Biotechnol.*, vol. 16, no. 1, pp. 1–12, 2016.
- [128] G. F. Pierce *et al.*, "Tissue repair processes in healing chronic pressure ulcers treated with recombinant platelet-derived growth factor BB," *Amer. J. Pathol.*, vol. 145, no. 6, p. 1399, 1994.
- [129] A. D. R. Li, K. B. Putra, L. Chen, J. S. Montgomery, and A. Shih, "Mosquito proboscis-inspired needle insertion to reduce tissue deformation and organ displacement," *Sci. Rep. (Nat. Publ. Group)*, vol. 10, no. 1, 2020, Art. no. 12248.
- [130] J. Margalit, R. Galun, and M. J. Rice, "Mouthpart sensilla of the tsetse fly and their function. I. Feeding patterns," *Ann. Trop. Med. Parasitol.*, vol. 66, no. 4, pp. 525–536, 1972.
- [131] M. J. Rice, R. Galun, and J. Margalit, "Mouthpart sensilla of the tsetse fly and their function. II. Labial sensilla," *Ann. Trop. Med. Parasitol.*, vol. 67, no. 1, pp. 101–107, 1973.
- [132] R. A. Leopold, T. P. Freeman, J. S. Buckner, and D. R. Nelson, "Mouthpart morphology and stylet penetration of host plants by the glassy-winged sharpshooter, *Homalodisca coagulata*, (Homoptera: Cicadellidae)," *Arthropod Struct. Develop.*, vol. 32, nos. 2–3, pp. 189–199, 2003.
- [133] G. J. Ma, L. T. Shi, and C. W. Wu, "Biomechanical property of a natural microneedle: The caterpillar spine," *J. Med. Devices*, vol. 5, no. 3, 2011, Art. no. 034502.
- [134] Z.-L. Zhao, H.-P. Zhao, G.-J. Ma, C.-W. Wu, K. Yang, and X.-Q. Feng, "Structures, properties, and functions of the stings of honey bees and paper wasps: A comparative study," *Biol. Open*, vol. 4, no. 7, pp. 921–928, 2015.
- [135] N. S. Al-Waili, "Topical application of natural honey, beeswax and olive oil mixture for atopic dermatitis or psoriasis: partially controlled, single-blinded study," *Complement. Ther. Med.*, vol. 11, no. 4, pp. 226–234, 2003.
- [136] D. B. Georgiev, A. R. Goudev, N. Manassiev, M. Metka, and J. C. Huber, "Effects of an herbal medication containing bee products on menopausal symptoms and cardiovascular risk markers: Results of a pilot open-uncontrolled trial," *Medscape Gen. Med.*, vol. 6, no. 4, p. 46, 2004.
- [137] R. D. Altman, D. R. Schultz, B. Collins-Yudiskas, J. Aldrich, P. I. Arnold, and H. E. Brown, "The effects of a partially purified fraction of an ant venom in rheumatoid arthritis," *Arthritis Rheumatism Off. J. Amer. Coll. Rheumatol.*, vol. 27, no. 3, pp. 277–284, 1984.
- [138] L. Moed, T. A. Shwayder, and M. W. Chang, "Cantharidin revisited: A blistering defense of an ancient medicine," *Archives Dermatol.*, vol. 137, no. 10, pp. 1357–1360, 2001.
- [139] D. A. Burns, "'Warts and all'—the history and folklore of warts: A review," *J. Roy. Soc. Med.*, vol. 85, no. 1, p. 37, 1992.
- [140] Z. Shi *et al.*, "A systematic review and meta-analysis of traditional insect Chinese medicines combined chemotherapy for non-surgical hepatocellular carcinoma therapy," *Sci. Rep.*, vol. 7, no. 1, pp. 1–24, 2017.
- [141] M. Zhu, X. Liu, C. Zhou, and J. Li, "Effect of sodium cantharidinate/vitamin B6 injection on survival, liver function, immune function, and quality of life in patients with hepatocellular carcinoma: Protocol for a meta-analysis," *Medicine*, vol. 99, no. 34, 2020, Art. no. e21952.
- [142] J.-H. Song, J.-M. Cha, B. C. Moon, W. J. Kim, S. Yang, and G. Choi, "Mantidis *Oötheca* (mantis egg case) original species identification via morphological analysis and DNA barcoding," *J. Ethnopharmacol.*, vol. 252, Apr. 2020, Art. no. 112574.
- [143] A. Jaggessar, H. Shahali, A. Mathew, and P. K. D. V. Yarlagaadda, "Biomimicking nano and micro-structured surface fabrication for antibacterial properties in medical implants," *J. Nanobiotechnol.*, vol. 15, no. 1, p. 64, 2017.
- [144] R. Ishwarya *et al.*, "Eco-friendly fabrication of Ag nanostructures using the seed extract of *Petalium murex*, an ancient Indian medicinal plant: Histopathological effects on the Zika virus vector *Aedes aegypti* and inhibition of biofilm-forming pathogenic bacteria," *J. Photochem. Photobiol. B, Biol.*, vol. 174, pp. 133–143, Sep. 2017.
- [145] M. A. Stewart, "The role of *Lucilia sericata* Meig. Larvae in osteomyelitis wounds," *Ann. Trop. Med. Parasitol.*, vol. 28, no. 4, pp. 445–460, 1934.
- [146] L. K. Ferguson and C. W. McLaughlin, Jr., "Maggot therapy: A rapid method of removing necrotic tissues," *Amer. J. Surg.*, vol. 29, no. 1, pp. 72–84, 1935.